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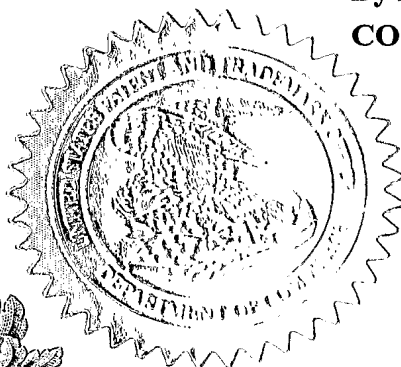
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COMPOSITIONS AND METHODS FOR TREATING PANCREATIC CANCER	
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Respectfully submitted,

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## DESCRIPTION

### COMPOSITIONS AND METHODS FOR TREATING PANCREATIC CANCER

#### 5 Technical Field

The present invention relates to the field of biological science, more specifically to the field of cancer research. In particular, the present invention relates a composition comprising a small interfering RNA (siRNA) of PCDH1, CDH3, GPR107 or EphA4.

#### 10 Background Art

Pancreatic ductal adenocarcinoma (PDACa) is the fifth leading cause of cancer death in the western world and has one of the highest mortality rates of any malignancy, with a 5-year survival rate only 4%. In USA, each year, estimated 30,700 patients are diagnosed with pancreatic cancer and nearly 30,000 will die of these diseases. The vast majority of patients are diagnosed at an advanced stage of disease at which it has no response to current therapies and the patients can survive for few months. Only surgical resection can offer the possibility of cure, but only 10-20% of patients with PDACa can undergo potentially curative resection and even after curative surgery, 80-90% of the patients relapse and die of the disease. Some improvements in surgical outcome or quality of life occur in patients who also receive chemotherapy including gemcitabine and/or radiation, although the impact on long-term survival has been minimal due to the intense resistance of PDACa to any treatment. At this point, management of most patients focuses on palliation.

Therefore, establishment of a novel molecular therapy for PDACa and identification of novel therapeutic molecular targets for PDACa are urgent issues for pancreatic cancer treatment now.

#### Disclosure of the Invention

30 The present invention based on the surprising discovery that small interfering RNAs (siRNAs) selective for PCDH1, CDH3, GPR107 or EphA4 are effective for

inhibiting the cellular growth of various cancer cells, including those involved in PDACa. The inventions described in this application are based in part on this discovery.

5 The invention provides methods for inhibiting cell growth. Among the methods provided are those comprising contacting a cell with a composition comprising a small interfering RNA (siRNA) of PCDH1, CDH3, GPR107 or EphA4. The invention also provides methods for inhibiting tumor cell growth in a subject. Such methods include administering to a subject a composition comprising a small interfering RNA (siRNA) of PCDH1, CDH3, GPR107 or EphA4. Another aspect of the invention provides methods for inhibiting the expression of the PCDH1, CDH3, GPR107 or EphA4 gene in a cell of a  
10 biological sample. Expression of the gene may be inhibited by introduction of a double stranded ribonucleic acid (RNA) molecule into the cell in an amount sufficient to inhibit expression of the PCDH1, CDH3, GPR107 or EphA4 gene. Another aspect of the invention relates to products including nucleic acid sequences and vectors as well as to compositions comprising them, useful, for example, in the provided methods. Among the  
15 products provided are siRNA molecules having the property to inhibit expression of the PCDH1, CDH3, GPR107 or EphA4 gene when introduced into a cell expressing said gene. Among such molecules are those that comprise a sense strand and an antisense strand, wherein the sense strand comprises a ribonucleotide sequence corresponding to a PCDH1, CDH3, GPR107 or EphA4 target sequence, and wherein the antisense strand comprises a  
20 ribonucleotide sequence which is complementary to said sense strand. The sense and the antisense strands of the molecule hybridize to each other to form a double-stranded molecule.

As used herein, the term "organism" refers to any living entity comprised of at least one cell. A living organism can be as simple as, for example, a single eukaryotic cell or as  
25 complex as a mammal, including a human being.

As used herein, the term "biological sample" refers to a whole organism or a subset of its tissues, cells or component parts (e.g. body fluids, including but not limited to blood, mucus, lymphatic fluid, synovial fluid, cerebrospinal fluid, saliva, amniotic fluid, amniotic cord blood, urine, vaginal fluid and semen). "Biological sample" further refers to a  
30 homogenate, lysate, extract, cell culture or tissue culture prepared from a whole organism or a subset of its cells, tissues or component parts, or a fraction or portion thereof. Lastly, "biological sample" refers to a medium, such as a nutrient broth or gel in which an

organism has been propagated, which contains cellular components, such as proteins or nucleic acid molecules.

The invention features methods of inhibiting cell growth. Cell growth is inhibited by contacting a cell with a composition of a small interfering RNA (siRNA) of PCDH1, CDH3, GPR107 or EphA4. The cell is further contacted with a transfection-enhancing agent. The cell is provided *in vitro*, *in vivo* or *ex vivo*. The subject is a mammal, *e.g.*, a human, non-human primate, mouse, rat, dog, cat, horse, or cow. The cell is a pancreatic ductal cell. Alternatively, the cell is a tumor cell (*i.e.*, cancer cell) such as a carcinoma cell or an adenocarcinoma cell. For example, the cell is a pancreatic ductal adenocarcinoma cell. By inhibiting cell growth is meant that the treated cell proliferates at a lower rate or has decreased viability than an untreated cell. Cell growth is measured by proliferation assays known in the art.

By the term "siRNA" is meant a double stranded RNA molecule which prevents translation of a target mRNA. Standard techniques of introducing siRNA into the cell are used, including those in which DNA is a template from which RNA is transcribed. The siRNA includes a sense PCDH1, CDH3, GPR107 or EphA4 nucleic acid sequence, an anti-sense PCDH1, CDH3, GPR107 or EphA4 nucleic acid sequence or both. The siRNA is constructed such that a single transcript has both the sense and complementary antisense sequences from the target gene, *e.g.*, a hairpin.

The method is used to alter gene expression in a cell in which expression of PCDH1, CDH3, GPR107 or EphA4 is upregulated, *e.g.*, as a result of malignant transformation of the cells. Binding of the siRNA to an PCDH1, CDH3, GPR107 or EphA4 transcript in the target cell results in a reduction in PCDH1, CDH3, GPR107 or EphA4 production by the cell. The length of the oligonucleotide is at least 10 nucleotides and may be as long as the naturally-occurring PCDH1, CDH3, GPR107 or EphA4 transcript. Preferably, the oligonucleotide is 19-25 nucleotides in length. Most preferably, the oligonucleotide is less than 75, 50, or 25 nucleotides in length. Examples of siRNA oligonucleotides of PCDH1, CDH3, GPR107 or EphA4 which inhibit PCDH1, CDH3, GPR107 or EphA4 expression in mammalian cells include oligonucleotides containing target sequences, for example, nucleotides of SEQ ID NOs: 54, 57, 60 or 66, respectively.

Methods for designing double stranded RNA having the ability to inhibit gene expression in a target cell are known. (See for example, US Patent No. 6,506,559, herein

incorporated by reference in its entirety). For example, a computer program for designing siRNAs is available from the Ambion website ([http://www.ambion.com/techlib/misc/siRNA\\_finder.html](http://www.ambion.com/techlib/misc/siRNA_finder.html)).

The computer program selects nucleotide sequences for siRNA synthesis based on the following protocol.

Selection of siRNA Target Sites

1. Beginning with the AUG start codon of the transcript, scan downstream for AA dinucleotide sequences. Record the occurrence of each AA and the 3' adjacent 19 nucleotides as potential siRNA target sites. Tuschl et al. recommend against designing siRNA to the 5' and 3' untranslated regions (UTRs) and regions near the start codon (within 75bases) as these may be richer in regulatory protein binding sites. UTR-binding proteins and/or translation initiation complexes may interfere with binding of the siRNA endonuclease complex.
2. Compare the potential target sites to the appropriate genome database (human, mouse, rat, etc.) and eliminate from consideration any target sequences with significant homology to other coding sequences. It is suggested to use BLAST, which can be found on the NCBI server at: [www.ncbi.nlm.nih.gov/BLAST/](http://www.ncbi.nlm.nih.gov/BLAST/)
3. Select qualifying target sequences for synthesis. Selecting several target sequences along the length of the gene to evaluate is typical.

Also included in the invention are isolated nucleic acid molecules that include the nucleic acid sequence of target sequences, for example, nucleotides of SEQ ID NOs: 54, 57, 60 and 66 or a nucleic acid molecule that is complementary to the nucleic acid sequence of nucleotides of SEQ ID NOs: 54, 57, 60 and 66. As used herein, an "isolated nucleic acid" is a nucleic acid removed from its original environment (e.g., the natural environment if naturally occurring) and thus, synthetically altered from its natural state. In the present invention, isolated nucleic acid includes DNA, RNA, and derivatives thereof. When the isolated nucleic acid is RNA or derivatives thereof, base "t" should be replaced with "u" in the nucleotide sequences. As used herein, the term "complementary" refers to Watson-Crick or Hoogsteen base pairing between nucleotides units of a nucleic acid molecule, and the term "binding" means the physical or chemical interaction between two polypeptides or compounds or associated polypeptides or compounds or combinations

thereof. Complementary nucleic acid sequences hybridize under appropriate conditions to form stable duplexes containing few or no mismatches. Furthermore, the sense strand and antisense strand of the isolated nucleotide of the present invention, can form double stranded nucleotide or hairpin loop structure by the hybridization. In a preferred

embodiment, such duplexes contain no more than 1 mismatch for every 10 matches. In an especially preferred embodiment, where the strands of the duplex are fully complementary, such duplexes contain no mismatches. The nucleic acid molecule is less than 3581, 3205, 6840 or 3468 nucleotides in length for PCDH1, CDH3, GPR107 or EphA4, respectively. For example, the nucleic acid molecule is less than 500, 200, or 75 nucleotides in length.

Also included in the invention is a vector containing one or more of the nucleic acids described herein, and a cell containing the vectors. The isolated nucleic acids of the present invention are useful for siRNA against PCDH1, CDH3, GPR107 or EphA4, or DNA encoding the siRNA. When the nucleic acids are used for siRNA or coding DNA thereof, the sense strand is preferably longer than 19 nucleotides, and more preferably longer than 21 nucleotides.

The invention is based in part on the discovery that the gene encoding PCDH1, CDH3, GPR107 or EphA4 is overexpressed in pancreatic ductal adenocarcinoma (PDACa) compared to non-cancerous pancreatic tissue. The cDNA of PCDH1, CDH3, GPR107 or EphA4 is 3581, 3205, 6840 or 3468 nucleotides in length. The nucleic acid and polypeptide sequences of PCDH1, CDH3, GPR107 or EphA4 are shown in SEQ ID NO: 1 and 2, 3 and 4, 5 and 6 or 7 and 8, respectively. The sequence data are also available via following accession numbers.

PCDH1(CFUPC): L11370, NM\_002587

CDH3: X63629, AB046844

GPR107: NM\_032925, (KIAA1624: R39794)

EphA4: L36645, NM\_004438

Transfection of siRNAs comprising SEQ ID NOs: 54, 57, 60 and 66 resulted in a growth inhibition of PDACa cell lines. PCDH1 (CFUPC) belongs to the protocadherin family, the largest subgroup of cadherin superfamily of calcium-dependent cell-cell adhesion molecules. Many of the protocadherin are highly expressed in the central nervous system and they are likely to play roles in neuronal circuit development and the modulation of synaptic transmission (Sano K, Tanihara H, Heimark RL, Obata S,



Davidson M, St John T, Taketani S, Suzuki S. Protocadherins: a large family of cadherin-related molecules in central nervous system. *EMBO J.*, 12:2249-56, 1993. Frank M, and Kemler R. Protocadherins. *Curr Opin Cell Biol.*, 14:557-62, 2002). However, PCDH1 is abundant in pancreatic cancer cells, but not in central nervous system (Figure 3A), and its function remains unknown.

CDH3 is also a classical member of the cadherin family (Shimoyama Y, Yoshida T, Terada M, Shimosato Y, Abe O, Hirohashi S. Molecular cloning of a human Ca<sup>2+</sup>-dependent cell-cell adhesion molecule homologous to mouse placental cadherin: its low expression in human placental tissues. *J Cell Biol.*, 109:1787-94. 1989) and they link to catenins and cytoskeletons through its conserved intracellular domain, mediating signal-transduction that control cell polarity, differentiation, motility and cell growth (Christofori G. Changing neighbours, changing behaviour: cell adhesion molecules-mediated signaling during tumor progression. *EMBO J.*, 22, 2318-2323, 2003). However, different from E-cadherin or N-cadherin, the function of CDH3 still remains unclear. Its expression is observed in mammary glands and ovary, and loss of expression was reported in breast cancer and prostate cancer, although the expression of P-cadherin in breast cancer correlates with poor prognosis (Peralta Soler A, Knudsen KA, Salazar H, Han AC, Keshgegian AA. P-cadherin expression in breast carcinoma indicates poor survival. *Cancer*, 86:1263-1272. 1999).

GPR107 (KIAA1624) is one of the G protein-coupled receptors (GPCR) with seven transmembranes. A large percentage of today's prescription drugs target one or more GPCRs with most major therapeutic area being served to some extent by several GPCR-based drugs. Clearly, GPCRs are in the highest rank in the terms of drug discovery potential. GPR107 is expressed unrestrictedly in normal heart, placenta, skeletal muscle, prostate, testis, ovary, spinal cord as shown in Northern blot analysis (Figure 3C). This is not abundant in major vital organs, suggesting that targeting for these molecules would be expected to lead less toxicity in human body.

EphA4 is one of the receptor with tyrosine kinase activity and their functions with their ephrin ligands are best studied in the nervous system, where Eph receptors and ephrin molecules are involved in patterning the developing hindbrain, axon pathfinding and guiding neural crest cell migration (Dodelet VC, and Pasquale EB. Eph receptors and ephrin ligands: embryogenesis to tumorigenesis. *Oncogene*, 19: 5614-5619, 2000). These

molecules also regulate embryonic vascular development and there are some reports about the association of Eph/ephrin with tumor angiogenesis (Dodelet VC, and Pasquale EB. Eph receptors and ephrin ligands: embryogenesis to tumorigenesis. *Oncogene*, 19: 5614-5619, 2000). The Eph receptor family consists of 13 members and their ligands, ephrins, are divided into two subclasses, the A-subclass (A1-A5) and the B-subclass (B1-B3). The receptors are divided on the basis of sequence similarity and ligand affinity into A-subclass (EphA1-A8), and B-subclass (EphB1-B4, B6). A-type receptors typically bind to most or all A-type ligands, and B-type receptors bind to most or all B-type ligands, with the exception of EphA4 that can bind both A-type and most B-type ligands (Dodelet VC, and Pasquale EB. Eph receptors and ephrin ligands: embryogenesis to tumorigenesis. *Oncogene*, 19: 5614-5619, 2000).

#### Methods of inhibiting cell growth

The present invention relates to inhibiting cell growth, i.e, cancer cell growth by inhibiting expression of PCDH1, CDH3, GPR107 or EphA4. Expression of PCDH1, CDH3, GPR107 or EphA4 is inhibited by small interfering RNA (siRNA) that specifically target the PCDH1, CDH3, GPR107 or EphA4 gene. PCDH1, CDH3, GPR107 or EphA4 targets include, for example, nucleotides of SEQ ID NOs: 54, 57, 60 and 66.

In non-mammalian cells, double-stranded RNA (dsRNA) has been shown to exert a strong and specific silencing effect on gene expression, which is referred as RNA interference (RNAi) (1). dsRNA is processed into 20-23 nucleotides dsRNA called small interfering RNA (siRNA) by an enzyme containing RNase III motif. The siRNA specifically targets complementary mRNA with a multicomponent nuclease complex (2, 3). In mammalian cells, siRNA composed of 20 or 21-mer dsRNA with 19 complementary nucleotides and 3' terminal noncomplementary dimmers of thymidine or uridine, have been shown to have a gene specific knock-down effect without inducing global changes in gene expression (4). In addition, plasmids containing small nuclear RNA (snRNA) U6 or polymerase III H1-RNA promoter effectively produce such short RNA recruiting type III class of RNA polymerase III and thus can constitutively suppress its target mRNA (5, 6).

The growth of cells are inhibited by contacting a cell, with a composition containing a siRNA of PCDH1, CDH3, GPR107 or EphA4. The cell is further contacted with a transfection agent. Suitable transfection agents are known in the art. By inhibition

of cell growth is meant the cell proliferates at a lower rate or has decreased viability compared to a cell not exposed to the composition. Cell growth is measured by methods known in the art such as, the MTT cell proliferation assay.

5 The siRNA of PCDH1, CDH3, GPR107 or EphA4 is directed to a single target of PCDH1, CDH3, GPR107 or EphA4 gene sequence. Alternatively, the siRNA is directed to multiple target of PCDH1, CDH3, GPR107 or EphA4 gene sequences. For example, the composition contains siRNA of PCDH1, CDH3, GPR107 or EphA4 directed to two, three, four, or five or more target sequences of PCDH1, CDH3, GPR107 or EphA4. By PCDH1, CDH3, GPR107 or EphA4 target sequence is meant a nucleotide sequence that is identical  
10 to a portion of the PCDH1, CDH3, GPR107 or EphA4 gene. The target sequence can include the 5' untranslated (UT) region, the open reading frame (ORF) or the 3' untranslated region of the human PCDH1, CDH3, GPR107 or EphA4 gene. Alternatively, the siRNA is a nucleic acid sequence complementary to an upstream or downstream modulator of PCDH1, CDH3, GPR107 or EphA4 gene expression. Examples of upstream  
15 and downstream modulators include, a transcription factor that binds the PCDH1, CDH3, GPR107 or EphA4 gene promoter, a kinase or phosphatase that interacts with the PCDH1, CDH3, GPR107 or EphA4 polypeptide, a PCDH1, CDH3, GPR107 or EphA4 promoter or enhancer.

siRNA of PCDH1, CDH3, GPR107 or EphA4 which hybridize to target mRNA  
20 decrease or inhibit production of the PCDH1, CDH3, GPR107 or EphA4 polypeptide product encoded by the PCDH1, CDH3, GPR107 or EphA4 gene by associating with the normally single-stranded mRNA transcript, thereby interfering with translation and thus, expression of the protein. The siRNA is less than 500, 200, 100, 50, or 25 nucleotides in length. Preferably the siRNA is 19-25 nucleotides in length. Exemplary nucleic acid  
25 sequence for the production of PCDH1, CDH3, GPR107 or EphA4 siRNA include the sequences of nucleotides of SEQ ID NOs: 54, 57, 60 or 66 as the target sequence, respectively. Furthermore, in order to enhance the inhibition activity of the siRNA, nucleotide "u" can be added to 3' end of the antisense strand of the target sequence. The number of "u"s to be added is at least 2, generally 2 to 10, preferably 2 to 5. The added  
30 "u"s form single strand at the 3' end of the antisense strand of the siRNA.

The cell is any cell that expresses or over-expresses PCDH1, CDH3, GPR107 or EphA4. The cell is an epithelial cell such as a pancreatic ductal cell. Alternatively, the

cell is a tumor cell such as a carcinoma, adenocarcinoma, blastoma, leukemia, myeloma, or sarcoma. The cell is a pancreatic ductal adenocarcinoma.

An siRNA of PCDH1, CDH3, GPR107 or EphA4 is directly introduced into the cells in a form that is capable of binding to the mRNA transcripts. Alternatively, the DNA  
5 encoding the siRNA of PCDH1, CDH3, GPR107 or EphA4 is in a vector.

Vectors are produced for example by cloning a PCDH1, CDH3, GPR107 or EphA4 target sequence into an expression vector operatively-linked regulatory sequences flanking the PCDH1, CDH3, GPR107 or EphA4 sequence in a manner that allows for expression (by transcription of the DNA molecule) of both strands (Lee, N.S., Dohjima, T., Bauer, G.,  
10 Li, H., Li, M.-J., Ehsani, A., Salvaterra, P., and Rossi, J. (2002) Expression of small interfering RNAs targeted against HIV-1 rev transcripts in human cells. Nature Biotechnology 20 : 500-505.). An RNA molecule that is antisense to PCDH1, CDH3, GPR107 or EphA4 mRNA is transcribed by a first promoter (e.g., a promoter sequence 3' of the cloned DNA) and an RNA molecule that is the sense strand for the PCDH1, CDH3,  
15 GPR107 or EphA4 mRNA is transcribed by a second promoter (e.g., a promoter sequence 5' of the cloned DNA). The sense and antisense strands hybridize *in vivo* to generate siRNA constructs for silencing of the PCDH1, CDH3, GPR107 or EphA4 gene. Alternatively, two constructs are utilized to create the sense and anti-sense strands of a siRNA construct. Cloned PCDH1, CDH3, GPR107 or EphA4 can encode a construct  
20 having secondary structure, e.g., hairpins, wherein a single transcript has both the sense and complementary antisense sequences from the target gene.

A loop sequence consisting of an arbitrary nucleotide sequence can be located between the sense and antisense sequence in order to form the hairpin loop structure. Thus, the present invention also provides siRNA having the general formula 5'-[A]-[B]-[A']-3',  
25 wherein [A] is a ribonucleotide sequence corresponding to a sequence selected from the group consisting of nucleotides of SEQ ID NOs: 54, 57, 60 and 66,

[B] is a ribonucleotide sequence consisting of 3 to 23 nucleotides, and

[A'] is a ribonucleotide sequence consisting of the complementary sequence of [A]

The region [A] hybridizes to [A'], and then a loop consisting of region [B] is  
30 formed. The loop sequence may be preferably 3 to 23 nucleotide in length. The loop sequence, for example, can be selected from group consisting of following sequences ([http://www.ambion.com/techlib/tb/tb\\_506.html](http://www.ambion.com/techlib/tb/tb_506.html)). Furthermore, loop sequence consisting

of 23 nucleotides also provides active siRNA (Jacque, J.-M., Triques, K., and Stevenson, M. (2002) Modulation of HIV-1 replication by RNA interference. *Nature* 418 : 435-438.).

CCC, CCACC or CCACACC: Jacque, J. M., Triques, K., and Stevenson, M (2002)

5 Modulation of HIV-1 replication by RNA interference. *Nature*, Vol. 418: 435-438.

UUCG: Lee, N.S., Dohjima, T., Bauer, G., Li, H., Li, M.-J., Ehsani, A., Salvaterra, P., and Rossi, J. (2002) Expression of small interfering RNAs targeted against HIV-1 rev transcripts in human cells. *Nature Biotechnology* 20 : 500-505. Fruscoloni, P., Zamboni, M., and Tocchini-Valentini, G. P. (2003) Exonucleolytic degradation of double-stranded

10 RNA by an activity in *Xenopus laevis* germinal vesicles. *Proc. Natl. Acad. Sci. USA* 100(4): 1639-1644..

UUCAAGAGA: Dykxhoorn, D. M., Novina, C. D., and Sharp, P. A. (2002) Killing the messenger: Short RNAs that silence gene expression. *Nature Reviews Molecular Cell Biology* 4: 457-467.

15

For example, preferable siRNAs having hairpin loop structure of the present invention are shown below. In the following structure, the loop sequence can be selected from group consisting of CCC, UUCG, CCACC, CCACACC, and UUCAAGAGA. Preferable loop sequence is UUCAAGAGA ("ttcaagaga" in DNA).

20

GACAUCAAUGACAACACAC-[B]-GUGUGUUGUCAUUGAUGUC (for target sequence of SEQ ID NO:54)

GGAGACAGGCUGGUUGUUG-[B]-CAACAACCAGCCUGUCUCC (for target sequence of SEQ ID NO:57)

25

GUGGCUCUACCAGCUCCUG-[B]-CAGGAGCUGGUAGAGCCAC (for target sequence of SEQ ID NO:60)

GCAGCACCAUCAUCCAUUG-[B]-CAAUGGAUGAUGGUGCUGC (for target sequence of SEQ ID NO:66)

30

The regulatory sequences flanking the PCDH1, CDH3, GPR107 or EphA4 sequence are identical or are different, such that their expression can be modulated independently, or in a temporal or spatial manner. siRNAs are transcribed intracellularly

by cloning the PCDH1, CDH3, GPR107 or EphA4 gene templates into a vector containing, e.g., a RNA pol III transcription unit from the small nuclear RNA (snRNA) U6 or the human H1 RNA promoter. For introducing the vector into the cell, transfection-enhancing agent can be used. FuGENE (Rochediagnostics), Lipofectamin 2000 (Invitrogen),  
5 Oligofectamin (Invitrogen), and Nucleofactor (Wako pure Chemical) are useful as the transfection-enhancing agent.

Oligonucleotides and oligonucleotides complementary to various portions of PCDH1, CDH3, GPR107 or EphA4 mRNA were tested *in vitro* for their ability to decrease production of PCDH1, CDH3, GPR107 or EphA4 in tumor cells (e.g., using the pancreatic  
10 cell line such as pancreatic ductal adenocarcinoma(PDACA) cell line) according to standard methods. A reduction in PCDH1, CDH3, GPR107 or EphA4 gene product in cells contacted with the candidate siRNA composition compared to cells cultured in the absence of the candidate composition is detected using specific antibodies of PCDH1, CDH3, GPR107 or EphA4 or other detection strategies. Sequences which decrease  
15 production of PCDH1, CDH3, GPR107 or EphA4 in *in vitro* cell-based or cell-free assays are then tested for their inhibitory effects on cell growth. Sequences which inhibit cell growth *in vitro* cell-based assay are test *in vivo* in rats or mice to confirm decreased PCDH1, CDH3, GPR107 or EphA4 production and decreased tumor cell growth in animals with malignant neoplasms.

#### Methods of treating malignant tumors

Patients with tumors characterized as over-expressing PCDH1, CDH3, GPR107 or EphA4 are treated by administering siRNA of PCDH1, CDH3, GPR107 or EphA4. siRNA  
25 therapy is used to inhibit expression of PCDH1, CDH3, GPR107 or EphA4 in patients suffering from or at risk of developing, for example, pancreatic ductal adenocarcinoma (PDACA). Such patients are identified by standard methods of the particular tumor type. Pancreatic ductal adenocarcinoma (PDACA) is diagnosed for example, by CT, MRI, ERCP, MRCP, computer tomography, or ultrasound. Treatment is efficacious if the treatment  
30 leads to clinical benefit such as, a reduction in expression of PCDH1, CDH3, GPR107 or EphA4, or a decrease in size, prevalence, or metastatic potential of the tumor in the subject. When treatment is applied prophylactically, "efficacious" means that the treatment retards or prevents tumors from forming or prevents or alleviates a symptom of clinical symptom.

of the tumor. Efficaciousness is determined in association with any known method for diagnosing or treating the particular tumor type.

siRNA therapy is carried out by administering to a patient a siRNA by standard vectors and/or gene delivery systems. Suitable gene delivery systems may include liposomes, receptor-mediated delivery systems, or viral vectors such as herpes viruses, retroviruses, adenoviruses and adeno-associated viruses, among others. A therapeutic nucleic acid composition is formulated in a pharmaceutically acceptable carrier. The therapeutic composition may also include a gene delivery system as described above. Pharmaceutically acceptable carriers are biologically compatible vehicles which are suitable for administration to an animal, e.g., physiological saline. A therapeutically effective amount of a compound is an amount which is capable of producing a medically desirable result such as reduced production of a PCDH1, CDH3, GPR107 or EphA4 gene product, reduction of cell growth, e.g., proliferation, or a reduction in tumor growth in a treated animal.

Parenteral administration, such as intravenous, subcutaneous, intramuscular, and intraperitoneal delivery routes, may be used to deliver siRNA compositions of PCDH1, CDH3, GPR107 or EphA4. For treatment of pancreatic tumors, direct infusion the celiac artery, splenic artery, or common hepatic artery, is useful.

Dosages for any one patient depends upon many factors, including the patient's size, body surface area, age, the particular nucleic acid to be administered, sex, time and route of administration, general health, and other drugs being administered concurrently. Dosage for intravenous administration of nucleic acids is from approximately  $10^6$  to  $10^{22}$  copies of the nucleic acid molecule.

The polynucleotides are administered by standard methods, such as by injection into the interstitial space of tissues such as muscles or skin, introduction into the circulation or into body cavities or by inhalation or insufflation. Polynucleotides are injected or otherwise delivered to the animal with a pharmaceutically acceptable liquid carrier, e.g., a liquid carrier, which is aqueous or partly aqueous. The polynucleotides are associated with a liposome (e.g., a cationic or anionic liposome). The polynucleotide includes genetic information necessary for expression by a target cell, such as a promoters.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this

invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

#### Brief Description of the Drawings

Figure 1 depicts photographs showing the results of validation of over expression of PCDH1 (A) and CDH3 (B) in the PDACa cells by RT-PCR. The microdissected normal pancreatic ductal epithelial cells (Normal) and vital organs (lung, heart, liver, kidney and bone marrow) from the same individual were compared by semiquantitative RT-PCR.

Figure 2 depicts photographs showing the result of immunohistochemistry in PDACa tissues. Overexpression of CDH3 and EphA4 protein was observed in pancreatic ductal adenocarcinoma, but not in normal pancreatic duct.

Figure 3 depicts photographs of Northern blot analysis showing the expression pattern in normal adult tissues of each target genes for pancreatic cancer. (A) PCDH1, (B) CDH3, (C) GPR107, and (D) EphA4.

Figure 4 depicts photographs showing the effect of Knocking-down endogenous PCDH1 in PDACa cell, PK-45P, by siRNA. Figure 4 (A) shows the results of RT-PCR. It validated knockdown effect of PCDH1 mRNA by transfection of siRNA expression vectors 410si, but not by 3344si, 3498si and EGFPsi. The 410si, 3344si, and 3498si were designed specifically for PCDH mRNA sequence, and EGFP was for EGFP mRNA sequence. RNA was harvested 48 hours after transfection and analyzed. ACTB was used to normalize input cDNA. Figure 4 (B) is a photograph showing the results of Colony formation assay. It showed drastic decrease of colony numbers in the cells one week after transfection with 410si that was validated to knock down PCDH1 effectively by RT-PCR. Figure 4 (C) is a photograph showing the results MTT assay. It also showed drastic decreased number of the grown cells transfected with 410si but not by 3344si, 3498si and EGFPsi.



Figure 5 depicts photographs showing the effect of Knocking-down endogenous CDH3 in PDACa cell, KLM-1, by siRNA. Figure 5 (A) shows the results of RT-PCR. It validated knockdown effect of CDH3 mRNA by transfection of siRNA expression vectors si24 but not by si29, si70 and EGFPsi. The si24, si29, and si70 were designed specifically for CDH3 mRNA sequence, and EGFPsi was for EGFP mRNA sequence. RNA was harvested 48 hours after transfection and analyzed. ACTB was used to normalize input cDNA. Figure 5 (B) is a photograph showing the results of Colony formation assay. It showed drastic decrease of colony numbers in the cells one week after transfection with si24 that was validated to knock down CDH3 effectively by RT-PCR. Figure 5 (C) is a photograph showing the results MTT assay. It also showed drastic decreased number of the grown cells transfected with si24, but not by si29, si70 and EGFPsi.

Figure 6 depicts photographs showing the effect of Knocking-down endogenous GPR107 in PDACa cell, KLM-1, by siRNA. Figure 6 (A) shows the results of RT-PCR. It validated knockdown effect of GPR107 mRNA by transfection of siRNA expression vectors 1003si, but not by 1066si, 1118si, 1171si and EGFPsi. The 1003si, 1066si, 1118si, and 1171si were designed specifically for GPR107 mRNA sequence, and EGFPsi was for EGFP mRNA sequence. RNA was harvested 48 hours after transfection and analyzed. ACTB was used to normalize input cDNA. Figure 6 (B) is a photograph showing the results of Colony formation assay. It showed decrease of colony numbers in the cells one week after transfection with 1003si that was validated to knock down GPR107 effectively by RT-PCR. Figure 6 (C) is a photograph showing the results MTT assay. It also showed decreased number of the grown cells transfected with 1003si, but not by 1118si, 1171si and EGFPsi.

Figure 7 depicts photographs showing the effect of Knocking-down endogenous EphA4 in PDACa cell, MIA-Paca2, by siRNA. Figure 7 (A) shows the results of RT-PCR. It validated knockdown effect of EphA4 mRNA by transfection of siRNA expression vectors 1313si, but not by 198si, 468si and EGFPsi. The 198si, 468si, 1313si were designed specifically for EphA4 mRNA sequence, and EGFPsi was for EGFP mRNA sequence. RNA was harvested 48 hours after transfection and analyzed. ACTB was used to normalize

input cDNA. Figure 7 (B) is a photograph showing the results of Colony formation assay. It showed drastic decrease of colony numbers in the cells one week after transfection with 1313si that was validated to knock down EphA4 effectively by RT-PCR. Figure 7 (C) is a photograph showing the results MTT assay. It also showed drastic decreased number of the grown cells transfected with 1313si, but not by 198si, 468si and EGFPsi.

#### Best Mode for Carrying out the Invention

The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

#### [Example 1] General Methods

##### *Cell lines and tissue specimens*

Human Pancreatic cell lines PK45P, KLM1 and MIA-PaCa2 (ATCC Number: CRL-1420) were obtained from the Cell Resource Center for Biomedical Research, Institute of Development, Aging and Cancer, Tohoku University. All these cells are publicly available.

##### *Isolation of over-expressing genes in PDACa cells by using cDNA microarray*

Fabrication of the cDNA microarray slides has been described (Ono K, Tanaka T, Tsunoda T, Kitahara O, Kihara C, Okamoto A, Ochiai K, Takagi T, and Nakamura Y. *Cancer Res.*, 60: 5007-5011, 2000). For each analysis of expression profiles it was prepared duplicate sets of cDNA microarray slides containing approximately 27,000 DNA spots, to reduce experimental fluctuation. Briefly, total RNA was purified from PDACa cells and normal pancreatic duct epithelium microdissected from 18 pancreatic cancer tissues. T7-based RNA amplification was carried out to obtain adequate RNA for microarray experiments. Aliquots of amplified RNA from PDACa cells and normal duct epithelium were labeled by reverse transcription with Cy5-dCTP and Cy3-dCTP, respectively (Amersham Biosciences). Hybridization, washing, and detection were carried out as described previously (Ono K, Tanaka T, Tsunoda T, Kitahara O, Kihara C, Okamoto A, Ochiai K, Takagi T, and Nakamura Y. *Cancer Res.*, 60: 5007-5011, 2000). Subsequently, among the up-regulated genes, it was focused four genes, PCDH1, CDH3, GPR107 and EphA4 because its expression ratio was greater than 5.0 in more than 50% of

informative cancers and their expression level in normal vital major organs was relatively low according to the our previous data of gene expression in 29 normal human tissues (Saito-Hisaminato A, Katagiri T, Kakiuchi S, Nakamura T, Tsunoda T, Nakamura Y. Genome-wide profiling of gene expression in 29 normal human tissues with a cDNA microarray. *DNA Res.*, 9: 35-45, 2002).

*Semiquantitative RT-PCR for PCDH1, CDH3, GPR107 and EphA4*

RNA from the microdissected PDACa cells and normal pancreatic ductal epithelial cells were subject to two-round amplification by T7-based *in vitro* transcription (Epicentre Technologies) and synthesized to single-strand cDNA. It was prepared appropriate dilutions of each single-stranded cDNA for subsequent PCR amplification by monitoring  $\beta$ -actin (ACTB) as a quantitative control. The primer sequences the present inventors used were 5'-AGAAGGAGACCAAGGACCTGTAT-3' (SEQ.ID.NO.9) and

5'-AGAACTTTATTGTCAGGGTCAAGG-3' (SEQ.ID.NO.10) for PCDH1,

5'-CTGAAGGCGGCTAACACAGAC-3' (SEQ.ID.NO.11) and

5'-TACACGATTGTCCTCACCTTC-3' (SEQ.ID.NO.12) for CDH3, and

5'-CATCCACGAAACTACCTTCAACT-3' (SEQ.ID.NO.13) and

5'-TCTCCTTAGAGAGAAGTGGGGTG-3' (SEQ.ID.NO.14) for ACTB. All

reactions involved initial denaturation at 94°C for 2 min followed by 21 cycles (for ACTB) or 28-32 cycles (for PCDH1 and CDH3) at 94°C for 30 s, 58°C for 30 s, and 72°C for 1 min, on a GeneAmp PCR system 9700 (PE Applied Biosystems).

*Immunohistochemistry*

Formalin-fixed and paraffin-embedded PDACa sections were immunostained using a mouse anti-CDH3 monoclonal antibody (BD Transduction Laboratories) or a rabbit anti-EPHA4 (EphA4) polyclonal antibody (Santa Cruz Biotechnology) for CDH3 and EPHA4 expression. Deparaffinized tissue sections were placed in 10 mM citrate buffer, pH 6.0, and heated to 108°C in an autoclave for 15 minutes for antigen retrieval. Sections were incubated with a 1:10 dilution or a 1:100 dilution of primary antibody for CDH3 or EPHA4, respectively, in a humidity chamber for an hour at room temperature, and developed with peroxidase labeled-dextran polymer followed by diaminobenzidine

(DAKO Envision Plus System; DAKO Corporation, Carpinteria, CA). Sections were counterstained with hematoxylin. For negative controls, primary antibody was omitted.

#### *Northern blot analysis*

- 5 Human multiple-tissue Northern blots (Clontech) were hybridized with a [ $\alpha$ -<sup>32</sup>P] dCTP-labeled PCR product amplified by the primers described above. Pre-hybridization, hybridization and washing were performed according to the supplier's recommendations. The blots were auto-radiographed with intensifying screens at -80°C for 5 days.

#### 10 Construction of psiU6BX Plasmid

The DNA fragment encoding siRNA was inserted into the GAP at nucleotide 485-490 as indicated (-) in the following plasmid sequence (SEQ ID No: 67).

```
GACGGATCGGGAGATCTCCCGATCCCCTATGGTGCACCTCTCAGTACAATCTGCTCTGGAT
CCACTAGTAACGGCCGCCAGTGTGCTGGAATTCTGGCTTGGGGATCAGCGTTTGAGTAAGA
GCCCCGCGTCTGAACCCCTCCGCGCCGCCCCGGCCCCAGTGGAAAGACGCGCAGGCAAAACG
CACCACGTGACGGAGCGTGACCGCGCGCCGAGCGCGCGCCAAGGTCGGGCAGGAAGAGGG
CCTATTTCCCATGATTCCCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAGATAAT
TAGAATTAATTTGACTGTAAACACAAAGATATTAGTACAAAATACGTGACGTAGAAAGTA
ATAATTTCTTGGGTAGTTTGCAGTTTTAAAAATTATGTTTTAAAAATGGACTATCATATGCT
TACCGTAACTTGAAAGTATTTTCGATTTCTTGGCTTTATATATCTTGTGGAAAGGACGAAA
CACC-----TTTTTACATCAGGTTGTTTTTCTGTTTGGTTTTTTTTTTTACACCACGTTT
ATACGCCGGTGCACGGTTTACCACCTGAAAACACC'TTTCATCTACAGGTGATATCTTTTAA
CACAAATAAAATGTAGTAGTCCTAGGAGACGGAATAGAAGGAGGTGGGGCCTAAAGCCGA
ATTCTGCAGATATCCATCACACTGGCGGCCGCTCGAGTGAGGCGGAAAGAACCAGCTGGG
GCTCTAGGGGGTATCCCCACGCGCCCTGTAGCGGCGCATTAAAGCGCGGCGGGTGTGGTGG
TTACGCGCAGCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTTCT
TCCCTTCCTTTCTCGCCACGTTTCGCCGGCTTTCCCCGTCAAGCTCTAAATCGGGGGCTCC
CTTTAGGGTTCCGATTTAGTGCTTTACGGCACCTCGACCCCAAAAAACTTGATTAGGGTG
ATGGTTCACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTTCGCCCTTTGACGTTGGAGT
CCACGTTCTTTAATAGTGGACTCTTGTTCCAAACTGGAACAACACTCAACCCATCTCGG
TCTATTCTTTTGATTTATAAGGGATTTTGCCGATTTTCGGCCTATTGGTTAAAAAATGAGC
TGATTTAACAAAAATTTAACGCGAATTAATCTGTGGAATGTGTGTCAGTTAGGGTGTGG
AAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGC
AACCAGGTGTGGAAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCT
CAATTAGTCAGCAACCATAGTCCCGCCCCCTAACTCCGCCCATCCCGCCCCCTAACTCCGCC
CAGTTCGCGCCATTCTCCGCCCCATGGCTGACTAATTTTTTTTTTATTATGCAGAGGCCGA
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GGCCGCCTCTGCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTTGGAGGCCTAGG  
CTTTTGC AAAAAGCTCCCGGGAGCTTGTATATCCATTTTCGGATCTGATCAAGAGACAGG  
ATGAGGATCGTTTCGCATGATTGAACAAGATGGATTGCACGCAGGTTCTCCGGCCGCTTG  
GGTGGAGAGGCTATTCCGGCTATGACTGGGCACAACAGACAATCGGCTGCTCTGATGCCGC  
CGTGTTCGGCTGTACGCGCAGGGGCGCCCGGTTCTTTTTGTCAAGACCGACCTGTCCGG  
TGCCCTGAATGAACTGCAGGACGAGGCAGCGCGGCTATCGTGGCTGGCCACGACGGGCGT  
TCCTTGCGCAGCTGTGCTCGACGTTGTCACTGAAGCGGGAAGGGACTGGCTGCTATTGGG  
CGAAGTGCCGGGGCAGGATCTCCTGTCTCATCTCACCTTGCTCCTGCCGAGAAAGTATCCAT  
CATGGCTGATGCAATGCGGCGGCTGCATACGCTTGATCCGGCTACCTGCCCATTCGACCA  
CCAAGCGAAACATCGCATCGAGCGAGCACGTACTCGGATGGAAGCCGGTCTTGTTCGATCA  
GGATGATCTGGACGAAGAGCATCAGGGGCTCGCGCCAGCCGAACGTTTCGCCAGGCTCAA  
GGCGCGCATGCCCGACGGCGAGGATCTCGTCGTGACCCATGGCGATGCCTGCTTGCCGAA  
TATCATGGTGGAAAATGGCCGCTTTTCTGGATTTCATCGACTGTGGCCGGCTGGGTGTGGC  
GGACCGCTATCAGGACATAGCGTTGGCTACCCGTGATATTGCTGAAGAGCTTGGCGGCGA  
ATGGGCTGACCGCTTCCTCGTGCTTTACGGTATCGCCGCTCCCGATTTCGCAGCGCATCGC  
CTTCTATCGCCTTCTTGACGAGTTCTTCTGAGCGGGACTCTGGGGTTTCAAATGACCGAC  
CAAGCGACGCCCCAACCTGCCATCACGAGATTTGATTCCACCGCCGCTTCTATGAAAGG  
TTGGGCTTCGGAATCGTTTTCCGGGACGCGGCTGGATGATCCTCCAGCGCGGGGATCTC  
ATGCTGGAGTTCTTCGCCCCACCCAACTTGTTTATTGCAGCTTATAATGGTTACAAATAA  
AGCAATAGCATCACAATTTTACAAATAAAGCATTTTTTTTCACTGCATTCTAGTTGTGGT  
TTGTCCAAACTCATCAATGTATCTTATCATGTCTGTATACCGTCGACCTCTAGCTAGAGC  
TTGGCGTAATCATGGTCATAGCTGTTTCTGTGTGAAATTGTTATCCGCTCACAATTCCA  
CACAACATACGAGCCGGAAGCATAAAGTGTAAGCCTGGGGTGCCTAATGAGTGAGCTAA  
CTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTCGTGCCAG  
CTGCATTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTGCCTATTGGGCGCTCTTCC  
GCTTCTCGCTCACTGACTCGCTGCGCTCGGTCGTTCCGGCTGCGGCGAGCGGTATCAGCT  
CACTCAAAGGCGGTAATACGTTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATG  
TGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTC  
CATAGGCTCCGCCCCCTGACGAGCATCACA AAAATCGACGCTCAAGTCAGAGGTGGCGA  
AACCAGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCT  
CCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTG  
GCGCTTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCCGGTGTAGGTCGTTCCGCTCCAAG  
CTGGGCTGTGTGCACGAACCCCCCGTTACGCCGACCGCTGCGCTTATCCGGTA ACTAT  
CGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTAAC  
AGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAAC  
TACGGCTACACTAGAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTC  
GGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGTTTTTTT  
GTTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTT  
TCTACGGGGTCTGACGCTCAGTGGAACGAAAACCTCACGTTAAGGGATTTTGGTCATGAGA

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TTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTAAAAATGAAGTTTAAATCAATC
TAAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCT
ATCTCAGCGATCTGTCTATTTCTGTTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATA
ACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCA
CGCTCACCGGCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGA
AGTGGTCTCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGA
GTAAGTAGTTTCGCCAGTTAATAGTTTTCGCAACGTTGTTGCCATTGCTACAGGCATCGTG
GTGTCACGCTCGTCGTTTGGTATGGCTTCATTCAGCTCCGGTCCCAACGATCAAGGCGA
GTTACATGATCCCCATGTTGTGCAAAAAGCGGTTAGCTCCTTCGGTCTCCGATCGTT
GTCAGAAGTAAGTTGGCCGAGTGTATCACTCATGGTTATGGCAGCACTGCATAATTCT
CTTACTGTTCATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCA
TTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATACGGGATAAT
ACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTGGAACGTTCTTCGGGGCGA
AAACTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCACTCGTGACCC
AACTGATCTTCAGCATCTTTTACTTTTACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGG
CAAAATGCCGCAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTTC
CTTTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGGATACATATTT
GAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCA
CCTGACGTC
```

snRNA U6 gene is reported to be transcribed by RNA polymerase III, which produce short transcripts with uridines at the 3' end. The genomic fragment of the snRNA U6 gene containing the promoter region was amplified by PCR using a set of primers,

5'-GGGGATCAGCGTTTGAGTAA-3' (SEQ ID No: 68), and

5 5'-TAGGCCCCACCTCCTTCTAT-3' (SEQ ID No: 69) and human placental DNA as a template. The product was purified and cloned into pCR plasmid vector using a TA cloning kit according to the supplier's protocol (Invitrogen). The *Bam*HI, *Xho*I fragment containing the snRNA U6 gene was purified and cloned into nucleotide 1257 to 56 fragment of pcDNA3.1(+) plasmid, which was amplified by PCR with a set of primer, 10 5'-TGCGGATCCAGAGCAGATTGTACTGAGAGT-3' (SEQ ID No: 70) and 5'-CTCTATCTCGAGTGAGGCGGAAAGAACCA-3' (SEQ ID No: 71). The ligated DNA was used for a template of PCR with primers, 5'-TTTAAGCTTGAAGACTATTTTACATCAGGTTGTTTTTCT-3' (SEQ ID No: 72) and

5'-TTTAAGCTTGAAGACACGGTGTTTCGTCCTTTCCACA-3' (SEQ ID No: 73). The product was digested with HindIII, which was subsequently self-ligated to produce psiU6BX vector plasmid. For the control, psiU6BX-BGFP was prepared by cloning double-stranded oligonucleotides of

5'- CACCGAAGCAGCACGACTTCTTCTTCAAGAGAGAAGAAGTCGTGCTGCTTC-3' (SEQ ID No: 74) and

5'- AAAAGAAGCAGCACGACTTCTTCTCTTGAAGAAGAAGTCGTGCTGCTTC -3' (SEQ ID No: 75) into the BbsI site in the psiU6BX vector.

#### 10 *siRNA-expressing constructs*

The nucleotide sequence of the siRNAs were designed using an siRNA design computer program available from the Ambion website. ([http://www.ambion.com/techlib/misc/siRNA\\_finder.html](http://www.ambion.com/techlib/misc/siRNA_finder.html)). Briefly, nucleotide sequences for siRNA synthesis are selected using the following protocol.

#### 15 *Selection of siRNA Target Sites:*

1. Starting with the AUG start codon of the each gene transcript, scan downstream for an AA dinucleotide sequences. The occurrence of each AA and the 3' adjacent 19 nucleotides are recorded as potential siRNA target sites. Tuschl et al. recommend against designing siRNA to the 5' and 3' untranslated regions (UTRs) and regions near the start codon (within 75bases) as these may be richer in regulatory protein binding sites. UTR-binding proteins and/or translation initiation complexes may interfere with binding of the siRNA endonuclease complex.

2. The potential target sites are compared to the appropriate genome database (human, mouse, rat, etc.) to eliminate target sequences with significant homology to other coding sequences.

3. Qualifying target sequences are selected for synthesis. Several target sequences along the length of the gene are selected for evaluation.

The oligonucleotides used for siRNAs of PCDH1, CDH3, GPR107 or EphA4 are shown below. Each oligonucleotide is a combination of a sense nucleotide sequence and an antisense nucleotide sequence of the target sequence. The nucleotide sequences of the hairpin loop structure and target sequence are shown in SEQ ID NO:54 to SEQ ID NO:57

and SEQ ID NO:60 to SEQ ID NO:66, respectively (endonuclease recognition sites are eliminated from each hairpin loop structure sequence).

Insert sequence of siRNA for PCDH1

5 410si:

5'-CACCGACATCAATGACAACACACTTCAAGAGAGTGTGTTGTCATTGATGTC-  
3' (SEQ ID NO: 15) and

5'-AAAAGACATCAATGACAACACACTCTCTTGAAGTGTGTTGTCATTGATGTC-  
3' (SEQ ID NO:16)

10

3344si:

5'-CACCGTCTACTCCAAACCTAGGTTTCAAGAGAACCTAGGTTTGGAGTAGAC-  
3' (SEQ ID NO: 17) and

15 5'-AAAAGTCTACTCCAAACCTAGGTTCTCTTGAACCTAGGTTTGGAGTAGAC-  
3' (SEQ ID NO: 18)

3498siRNA:

5'-CACCCCTCTTCCTCACCCTAGGTTCAAGAGACCTAGTGGTGAGGAAGAGG-  
3' (SEQ ID NO: 19) and

20 5'-AAAACCTCTTCCTCACCCTAGGTTCTCTTGAACCTAGTGGTGAGGAAGAGG-  
3' (SEQ ID NO: 20)

Insert sequence of siRNA for CDH3

si24:

25 5'-CACCGGAGACAGGCTGGTTGTTGTTCAAGAGACAACAACCAGCCTGTCTCC-  
3' (SEQ ID NO: 21)and

5'-AAAAGGAGACAGGCTGGTTGTTGTCTCTTGAACAACAACCAGCCTGTCTCC-  
3' (SEQ ID NO: 22)

30 si29:

5'-CACCCATCTCCATCATCGTGACCTTCAAGAGAGGTCACGATGATGGAGATG-  
3' (SEQ ID NO: 23)and



5'-AAAACATCTCCATCATCGTGACCTCTCTTGAAGGTCACGATGATGGAGATG-  
3' (SEQ ID NO: 24)

si70:

5 5'-CACCCATCACGGACAAGGACCTGTTCAAGAGACAGGTCCTTGTCCGTGATG-  
3' (SEQ ID NO: 25)and

5'-AAAACATCACGGACAAGGACCTGTCTCTTGAACAGGTCCTTGTCCGTGATG-  
3' (SEQ ID NO: 26)

10 Insert sequence of siRNA for GPR107

1003si:

5'-CACCGTGGCTCTACCAGCTCCTGTTCAAGAGACAGGAGCTGGTAGAGCCAC-  
3' (SEQ ID NO: 27)and

15 5'-AAAAGTGGCTCTACCAGCTCCTGTCTCTTGAACAGGAGCTGGTAGAGCCAC-  
3' (SEQ ID NO: 28)

1066si:

5'-CACCATTCCGTCCGGCTTCAGATTTCAGAGAATCTGAAGCCGGACGGAAT-  
3' (SEQ ID NO: 29)and

20 5'-AAAAATTCCGTCCGGCTTCAGATTCTCTTGAATCTGAAGCCGGACGGAAT-  
3' (SEQ ID NO: 30)

1118si:

25 5'-CACCGACTTGGAATGGAGTCCGTTCAAGAGACGGACTCCATTTCCAAGTC-  
3' (SEQ ID NO: 31)and

5'-AAAAGACTTGGAATGGAGTCCGTCTCTTGAACGGACTCCATTTCCAAGTC-  
3' (SEQ ID NO: 32)

1171si:

30 5'-CACCGAAAGTCAAGAAGGTGACCTTCAAGAGAGGTCACCTTCTTGACTTTC-  
3' (SEQ ID NO: 33) and

5'-AAAAGAAAGTCAAGAAGGTGACCTCTCTTGAAGGTCACCTTCTTGACTTTC-  
3' (SEQ ID NO: 34)

Insert sequence of siRNA for EphA4

5 198si:

5'-CACCTCCGAACCTACCAAGTGTGTTCAAGAGACACACTTGGTAGGTTTCGGA-  
3' (SEQ ID NO: 35)and

5'-AAAATCCGAACCTACCAAGTGTGTCTCTTGAACACACTTGGTAGGTTTCGGA-  
3' (SEQ ID NO: 36)

10

468si:

5'-CACCTCATGAAGCTGAACACCGATTCAAGAGATCGGTGTTTCAGCTTCATGA-  
3' (SEQ ID NO: 37)and

5'-AAAATCATGAAGCTGAACACCGATCTCTTGAATCGGTGTTTCAGCTTCATGA-  
15 3' (SEQ ID NO: 38)

1313si:

5'-CACCGCAGCACCATCATCCATTGTTCAAGAGACAATGGATGATGGTGCTGC-  
3' (SEQ ID NO: 39) and

20 5'-AAAAGCAGCACCATCATCCATTGTCTCTTGAACAATGGATGATGGTGCTGC-  
3' (SEQ ID NO: 40)

Insert sequence of siRNA for control

EGFPsi: (control)

25 5'- CACCGAAGCAGCACGACTTCTTCTTCAAGAGAGAAGAAGTCGTGCTGCTTC  
-3' (SEQ ID NO: 74) and

5'-AAAAGAAGCAGCACGACTTCTTCTCTTGAAGAAGAAGTCGTGCTGCTTC-  
3' (SEQ ID NO: 75)

Sequence ID NO of each sequences are listed in Table1

gene	siRNA	effect	insert seq SEQ ID NO		hairpin siRNA	target SEQ ID NO	position
PCDH1	410si	+	15	16	41	54	595-613
PCDH1	3344si	-	17	18	42	55	3565-3583
PCDH1	3498si	-	19	20	43	56	3719-3737
CDH3	si24	+	21	22	44	57	556-574
CDH3	si29	-	23	24	45	58	670-688
CDH3	si70	-	25	26	46	59	1768-1786
GPR107	1003si	+	27	28	47	60	1570-1588
GPR107	1066si	-	29	30	48	61	1633-1651
GPR107	1118si	-	31	32	49	62	1685-1703
GPR107	1171si	-	33	34	50	63	1738-1756
EphA4	198si	-	35	36	51	64	242-260
EphA4	468si	-	37	38	52	65	530-548
EphA4	1313si	+	39	40	53	66	1357-1375
control	EGFPsi	-	74	75			

#### colony formation / MTT assay

5 Human PDACa cell lines among PK45P, KLM1 and MIA-PaCa2, were plated onto 10-cm dishes ( $5 \times 10^5$  cells/dish) and transfected with psiU6BX containing EGFP target sequence (EGFP) and psiU6BX containing target sequence using Lipofectamine 2000 (Invitrogen) or FuGENE6 (Roche), according to manufacture's instruction. Cells were selected by 500 mg/ml Geneticin for one week, and preliminary cells were harvested 10 48 hours after transfection and analyzed by RT-PCR to validate knockdown effect on PCDH1, CDH3, GPR107 and EphA4. The primers of RT-PCR were the same ones described above. These cells were also stained by Giemsa solution and performed MTT assay to evaluate the colony formation and the cell number, respectively.

15 [Example 2] Reduction of the expression of the genes PCDH1, CDH3, GPR107 or EphA4 and growth suppression of cancer cells by siRNA

In previous study, it was generated precise expression profiles of PDACa by combining laser microdissection with genome-wide cDNA microarrays with 27,000 genes spotted. The present inventors identified more than 200 genes as up-regulated genes in 20 PDACa cells comparing with the expression pattern of normal pancreatic ductal epithelium that was thought to be the origin of PDACa (Nakamura T, Furukawa Y, Nakagawa H,

Tsunoda T, Ohgashi H, Murata K, Ishikawa O, Ohgaki, Kashimura N, Miyamoto M, Hirano S, Kondo S, Katoh H, Nakamura Y, and Katagiri T. Genome-wide cDNA microarray analysis of gene-expression profiles in pancreatic cancers using populations of tumor cells and normal ductal epithelium cells selected for purity by laser microdissection. *Oncogene*, 2004 Feb 9, Epub ahead of print). Based on these expression profile of PDACa cells, the present inventors selected four over-expressing genes, PCDH1 and CDH3 and validated their overexpression in PDACa by RT-PCR using the cDNA from microdissected PDACa cells (Figure 1A,B) or immunohistochemistry (Figure 2). Their products are supposed to be cell-surface membrane proteins that are ideal molecule target for drug design and antibody therapy against cancer. Clinical trials approved that Trastuzumab (Herceptin), a humanized monoclonal antibody against ERBB2 (Her2) is effective for subsets of metastatic breast cancer with HER2 over-expressed, and cell-surface molecules that mediates signaling process necessary for essential cellular functions and for maintaining the malignant phenotypes are now most promising targets for cancer therapy (Pegram M, and Slamon DJ. Biological rationale for Her2/neu as a target for monoclonal antibody therapy. *Semin.Oncology*, 27 (suppl 9): 13-19, 2000). Drug design targeting these membrane molecules can be approached both by blocking their growth-promoting signals and/or by modulating ADCC activity in the same way with Trastuzumab.

(1) PCDH1 (Protocadherin 1) (Genbank Accession No.NM\_002587; SEQ ID No.1,2)

To investigate the growth or survival effect of PCDH1 on PDACa cells, the present inventors knocked down their endogenous expression of PCDH1 specifically by mammalian vector-based RNA interference (RNAi) technique in PDACa cell line. PCDH1 is expressed unrestrictedly in normal heart, placenta, prostate as shown in Northern blot analysis (Figure 3A). This is not abundant in major vital organs, suggesting that targeting for these molecules would be expected to lead less toxicity in human body.

The transfection of the siRNA-producing vectors clearly resulted in reduction of the endogenous expression in one designed siRNA, 410si, for PCDH1 (Figure 4A). This knocking-down effect by the siRNA on PCDH1 mRNA resulted in drastic growth suppression in colony formation assay (Figure 4B) and MTT assay (Figure 4C). These findings strongly suggested that overexpression of PCDH1 in PDACa cells were associated with cancer cell viability. PCDH1 and other protocadherins are supported to have homophilic interaction on the cell surface by means of their cadherin domains and

modulate intercellular signal transduction for cytoskeleton conformation, cell motility or cell growth (Sano K, Tanihara H, Heimark RL, Obata S, Davidson M, St John T, Taketani S, Suzuki S. Protocadherins: a large family of cadherin-related molecules in central nervous system. *EMBO J.*, 12:2249-56, 1993, Frank M, and Kemler R. Protocadherins.

5 *Curr Opin Cell Biol.*, 14:557-62, 2002.). According to our data, PCDH1 is likely to modulate positive signal for pancreatic cancer cell growth through its homophilic interaction in cell-cell adhesion.

(2) CDH3 (P-cadherin) (Genbank Accession No.NM\_001793; SEQ ID No.3,4)

10 The present inventors validated CDH3 overexpression in PDACa cells by RT-PCR (Figure 1B) and immunohistochemistry (Figure 2A), and according to the microarray data and RT-PCR (Figure 1B), CDH3 overexpression was one of the most predominant patterns among more than 200 up-regulated genes in our PDACa profiles. CDH3 is expressed unrestrainedly in normal thymus, prostate, ovary, trachea as shown in Northern blot analysis (Figure 3B). This is not abundant in major vital organs, suggesting that targeting for these  
15 molecules would be expected to lead less toxicity in human body.

To investigate the growth or survival effect of CDH3 on PDACa cells, the present inventors knocked down their endogenous expression of CDH3 specifically by mammalian vector-based RNA interference (RNAi) technique in PDACa cell line. The transfection of the siRNA-producing vectors clearly resulted in reduction of the endogenous expression in  
20 one designed siRNA, si24, for CDH3 (Figure 5A). This knocking-down effect by the siRNA on CDH3 mRNA resulted in drastic growth suppression in colony formation assay (Figure 5B) and MTT assay (Figure 5C). These findings strongly suggested that overexpression of CDH3 in PDACa cells were associated with cancer cell viability as well as cell-cell interaction, and this molecule may involve signal transduction from cell-cell  
25 interaction. PDACa is extremely aggressive and high expression of CDH3 in PDACa may be associated with their aggressiveness and metastatic potential as well.

(3) GPR107 (G protein-coupled receptor 107) (Genbank Accession No. AB046844; SEQ ID No.5,6)

30 The present inventors identified this orphan GPCR as a target for pancreas cancer, which function and ligands are unknown. To investigate the growth or survival effect of GPR107 on PDACa cells, the present inventors knocked down their endogenous expression of GPR107 specifically by siRNA in PDACa cell line. The transfection of the

siRNA-producing vectors clearly resulted in reduction of the endogenous expression in one designed siRNA, 1003si, for GPR107 (Figure 6A). This knocking-down effect by the siRNA on GPR107 mRNA resulted in growth suppression in colony formation assay (Figure 6B) and MTT assay (Figure 6C). These findings strongly suggested that overexpression of GPR107 in PDACa cells were associated with cancer cell viability. Hence, these findings suggested that blocking by antibody or antagonist for GPR107 is a promising approach for PDACa treatment.

(4) EphA4 (Genbank Accession No.NM\_004438; SEQ ID No.7,8)

The present inventors validated EphA4 overexpression in PDACa by RT-PCR and immunohistochemistry (Figure 2B), but in pancreatic cancer tissues, the ligand of EphA4 is unknown. Northern blot analysis (Figure 3D) showed that EphA4 was abundant in testis, not in central nervous system and other major organs. Recently the antibody targeting against other Eph receptor family member, EphA2 that is also over-expressed in several cancers, was reported to inhibit breast cancer cell growth *in vitro* and *in vivo* (Carles-Kinch K, Kilpatrick KE, Stewart JC, Kinch MS. Antibody targeting of the EphA2 tyrosine kinase inhibits malignant cell behavior. *Cancer Res.*, 62:2840-2847, 2002). However, EphA2 is expressed ubiquitously in adult tissues, indicating much more possibility of toxicity in treatment of antibody therapy. To investigate the growth or survival effect of EphA4 on PDACa cells, the present inventors knocked down their endogenous expression of EphA4 specifically by siRNA in PDACa cell line. The transfection of the siRNA-producing vectors clearly resulted in reduction of the endogenous expression in one designed siRNA, 1313si, for EphA4 (Figure 7A). This knocking-down effect by the siRNA on EphA4 mRNA resulted in drastic growth suppression in colony formation assay (Figure 7B) and MTT assay (Figure 7C). Considering its tyrosine kinase activity, membrane localization and its specific expression pattern, EphA4 is one the most ideal molecular targets for pancreatic cancer.

In conclusion, the present inventors identified four membrane-type molecules over-expressed in PDACa cells and all of them are likely to be associated with cancer cell growth, suggested these membrane-type molecules are ideal molecular targets for deadly pancreatic cancer treatment and antibodies against these membrane molecules are promising therapeutic approach.

### Industrial Applicability

The present inventors have shown that the cell growth is suppressed by small interfering RNA (siRNA) that specifically target the PCDH1, CDH3, GPR107 or EphA4 gene. Thus, this novel siRNAs are useful target for the development of anti-cancer pharmaceuticals. For example, agents that block the expression of PCDH1, CDH3, GPR107 or EphA4 or prevent its activity may find therapeutic utility as anti-cancer agents, particularly anti-cancer agents for the treatment of pancreatic cancer, such as pancreatic ductal adenocarcinoma (PDACa).

While the invention has been described in detail and with reference to specific embodiments thereof, it will be apparent to one skilled in the art that various changes and modifications can be made therein without departing from the spirit and scope of the invention.

### References

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- (2) Hammond SM, Bernstein E, Beach D, Hannon GJ. An RNA-directed nuclease mediates post-transcriptional gene silencing in Drosophila cells. Nature. 2000 Mar 16;404(6775):293-6.
- (3) Hannon GJ. RNA interference. Nature. 2002 Jul 11;418(6894):244-51.
- (4) Elbashir SM, Harborth J, Lendeckel W, Yalcin A, Weber K, Tuschl T. Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells. Nature. 2001 May 24;411(6836):494-8.
- (5) Miyagishi M, Taira K. U6 promoter-driven siRNAs with four uridine 3' overhangs efficiently suppress targeted gene expression in mammalian cells. Nat Biotechnol. 2002 May;20(5):497-500.
- (6) Brummelkamp TR, Bernards R, Agami R. A System for Stable Expression of Short Interfering RNAs in Mammalian Cells Science. 296(5567):550-553, April 19, 2002.

CLAIMS

1. A method for treating or preventing pancreatic cancer in a subject comprising administering to said subject a composition comprising a small interfering RNA (siRNA) of *PCDH1*, *CDH3*, *GPR107* or *EPHA4*.
2. The method of claim 1, wherein said siRNA comprises a sense nucleic acid and an anti-sense nucleic acid of *PCDH1*, *CDH3*, *GPR107* or *EPHA4*.
3. The method of claim 1, wherein the pancreatic cancer is an pancreatic ductal adenocarcinoma (PDACa).
4. The method of claim 2, wherein the siRNA comprises a ribonucleotide sequence corresponding to a sequence selected from the group consisting of SEQ ID NOs: 54, 57, 60 and 66 as the target sequence.
5. The method of claim 4, said siRNA has the general formula 5'-[A]-[B]-[A']-3', wherein [A] is a ribonucleotide sequence corresponding to a sequence selected from the group consisting of nucleotides of SEQ ID NOs: 54, 57, 60 and 66. [B] is a ribonucleotide sequence consisting of 3 to 23 nucleotides, and [A'] is a ribonucleotide sequence consisting of the complementary sequence of [A].
6. The method of claim 1, wherein said composition comprises a transfection-enhancing agent.
7. A double-stranded molecule comprising a sense strand and an antisense strand, wherein the sense strand comprises a ribonucleotide sequence corresponding to a target sequence selected from the group consisting of SEQ ID NOs: 54, 57, 60 and 66, and wherein the antisense strand comprises a ribonucleotide sequence which is complementary to said sense strand, wherein said sense strand and said antisense strand hybridize to each other to form said double-stranded molecule, and wherein said double-stranded molecule, when introduced into a cell expressing the *PCDH1*, *CDH3*, *GPR107* or *EPHA4* gene, inhibits expression of said gene.
8. The double-stranded molecule of claim 7, wherein said target sequence comprises at least about 10 contiguous nucleotides from the nucleotide sequences selected from the group of SEQ ID NOs: 1, 3, 5, and 7.



9. The double-stranded molecule of claim 8, wherein said target sequence comprises from about 19 to about 25 contiguous nucleotides from the nucleotide sequences selected from the group of SEQ ID NOs: 1, 3, 5, and 7.
5. 10. The double-stranded molecule of claim 9, wherein said double-stranded molecule is a single ribonucleotide transcript comprising the sense strand and the antisense strand linked via a single-stranded ribonucleotide sequence.
11. The double-stranded molecule of claim 8, wherein the double-stranded molecule is an oligonucleotide of less than about 100 nucleotides in length.
10. 12. The double-stranded molecule of claim 11, wherein the double-stranded molecule is an oligonucleotide of less than about 75 nucleotides in length.
13. The double-stranded molecule of claim 12, wherein the double-stranded molecule is an oligonucleotide of less than about 50 nucleotides in length.
14. The double-stranded molecule of claim 13, wherein the double-stranded molecule is an oligonucleotide of less than about 25 nucleotides in length.
15. 15. The double-stranded polynucleotide of claim 14, wherein the double stranded molecule is an oligonucleotide of between about 19 and about 25 nucleotides in length.
16. A vector encoding the double-stranded molecule of claim 8.
20. 17. The vector of claim 16, wherein the vector encodes a transcript having a secondary structure and comprises the sense strand and the antisense strand.
18. The vector of claim 17, wherein the transcript further comprises a single-stranded ribonucleotide sequence linking said sense strand and said antisense strand.
25. 19. A vector comprising a polynucleotide comprising a combination of a sense strand nucleic acid and an antisense strand nucleic acid, wherein said sense strand nucleic acid comprises nucleotide sequence of SEQ ID NOs: 54, 57, 60 and 66, and said antisense strand nucleic acid consists of a sequence complementary to the sense strand.
20. The vector of claim 19, wherein said polynucleotide has the general formula  
5'-[A]-[B]-[A']-3'

wherein [A] is a nucleotide sequence of SEQ ID NOs: 54, 57, 60 and 66; [B] is a nucleotide sequence consisting of 3 to 23 nucleotides; and [A'] is a nucleotide sequence complementary to [A].

- 5 21. A pharmaceutical composition for treating or preventing pancreatic cancer comprising a pharmaceutically effective amount of a small interfering RNA (siRNA) of *PCDH1*, *CDH3*, *GPR107* or *EPHA4* as an active ingredient, and a pharmaceutically acceptable carrier..
- 10 22. The pharmaceutical composition of claim 21, wherein the siRNA comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 54, 57, 60 and 66 as the target sequence.
- 15 23. The composition of claim 22, wherein the siRNA has the general formula  
5'-[A]-[B]-[A']-3'  
wherein [A] is a ribonucleotide sequence corresponding to a nucleotide sequence of SEQ ID NOs: 54, 57, 60 and 66; [B] is a ribonucleotide sequence consisting of 3 to 23 nucleotides; and [A'] is a ribonucleotide sequence complementary to [A].

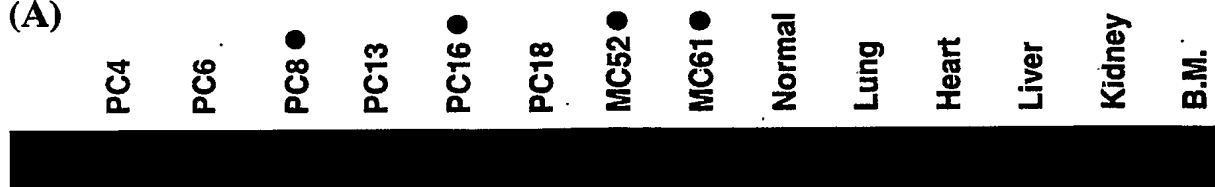
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ABSTRACT

5 The invention features a method for inhibiting growth of a cancer cell by contacting the cell with a composition of a siRNA of *PCDH1*, *CDH3*, *GPR107* or *EPHA4*. Methods of treating cancer are also within the invention. The invention also features products, including nucleic acid sequences and vectors as well as to compositions comprising them, useful in the provided methods. The invention also provides a method for inhibiting of tumor cell, for example pancreatic cancer cell, particularly pancreatic ductal adenocarcinoma (PDACa).

Fig: 1

(A)



(B)

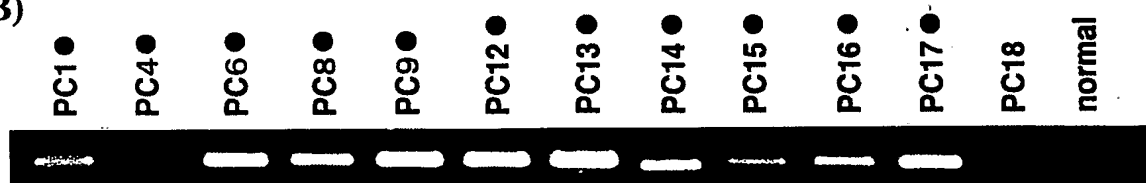
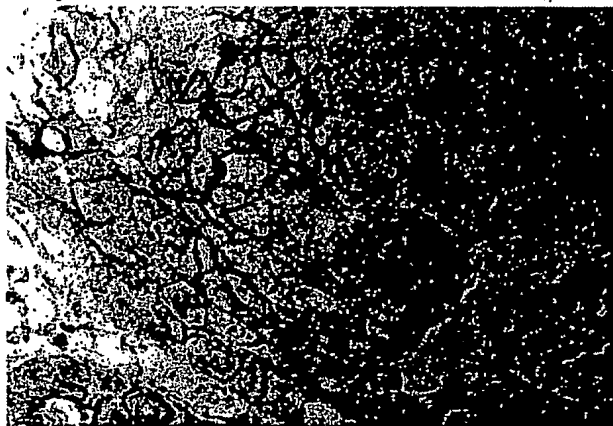


Fig. 2

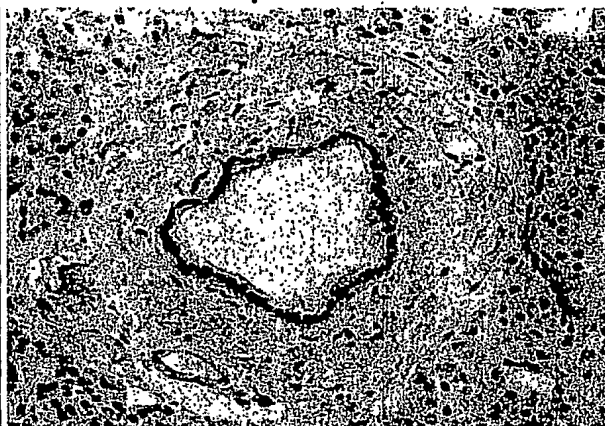
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**(A) CDH3**

**pancreatic ductal adenocarcinoma**

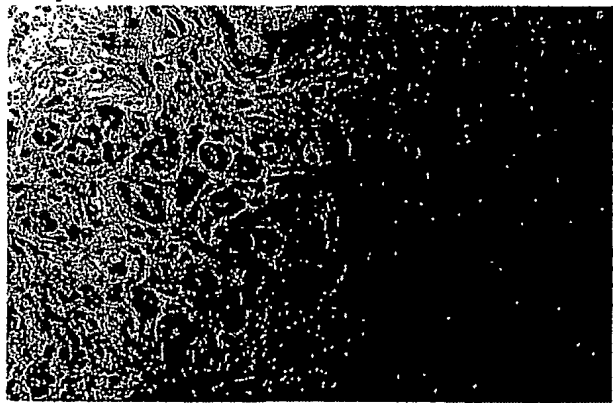


**normal pancreatic duct**



**(B) EphA4**

**pancreatic ductal adenocarcinoma**

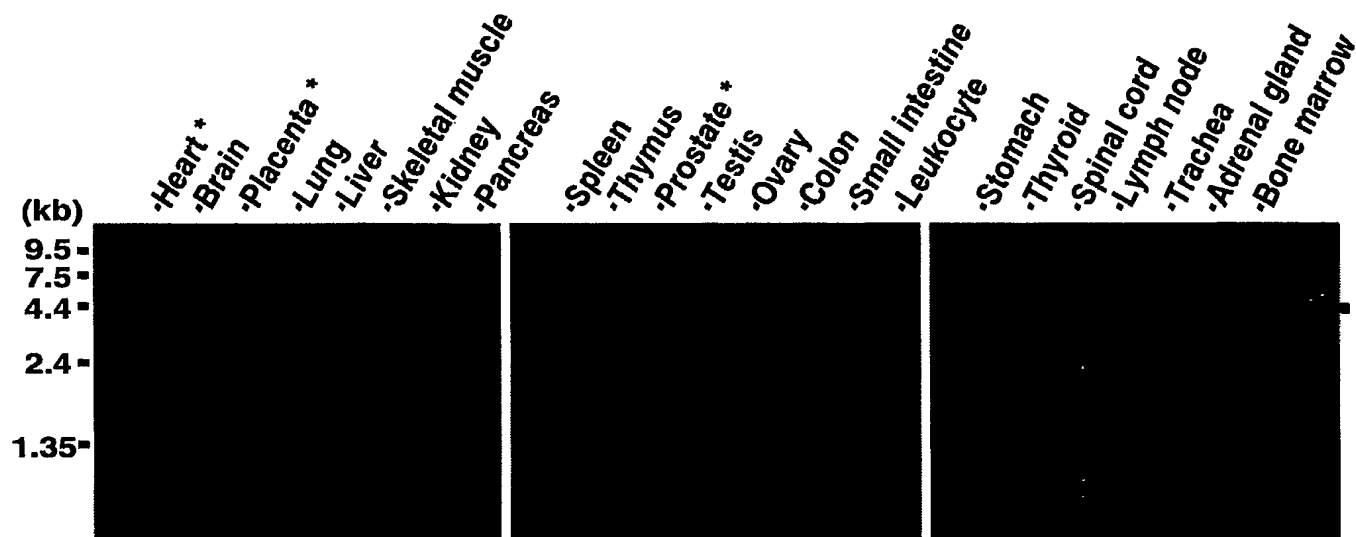
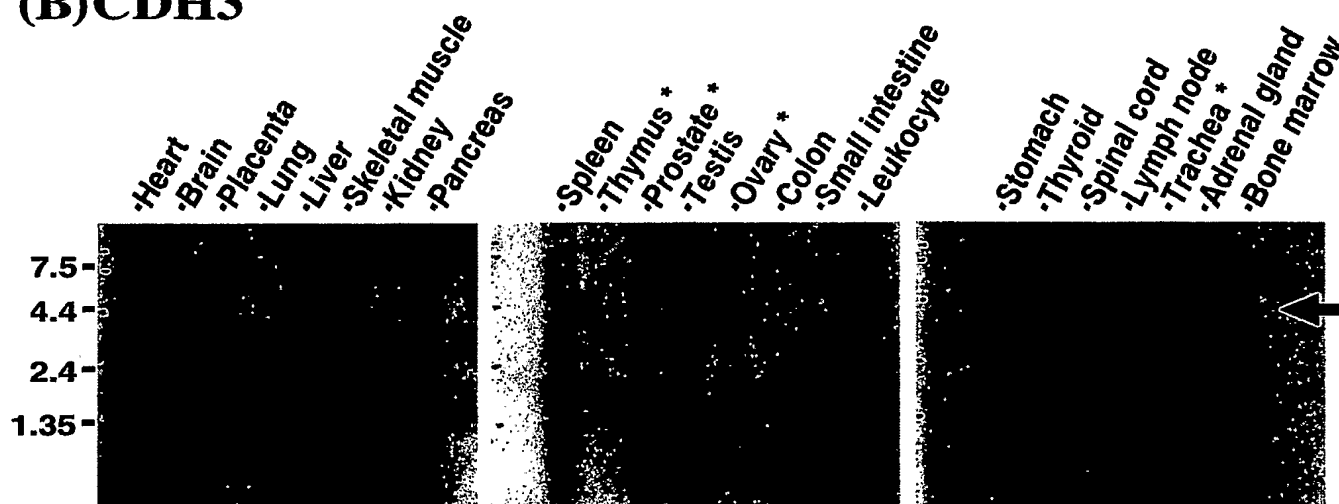


**normal pancreatic duct**



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Fig. 3-1

**(A) PCDH1****(B) CDH3**

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Fig. 3-2

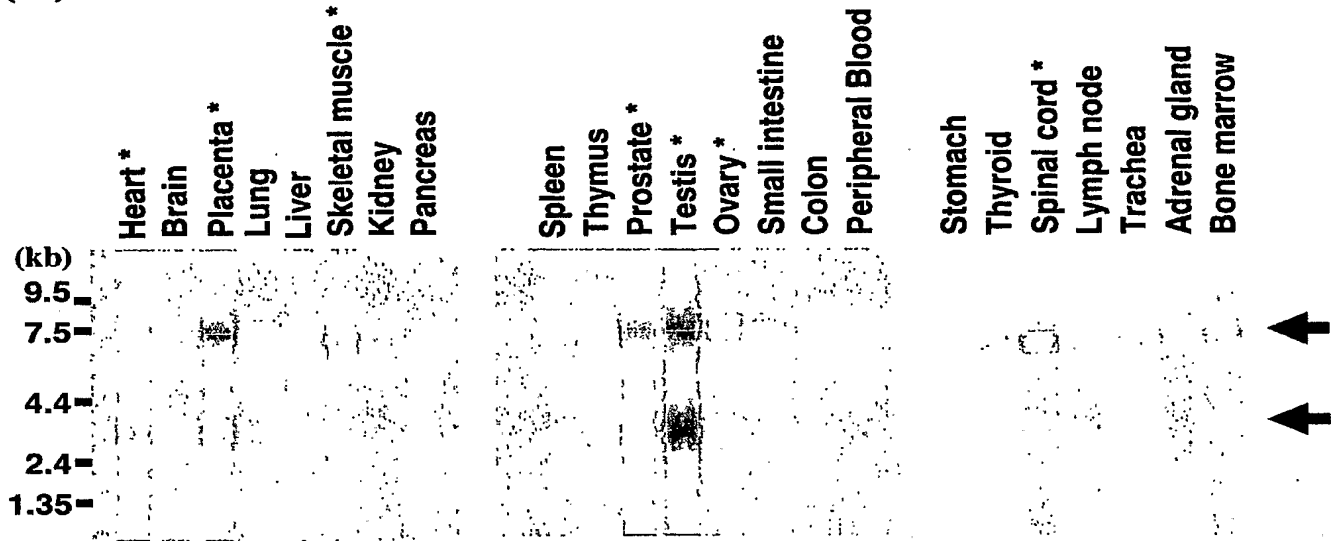
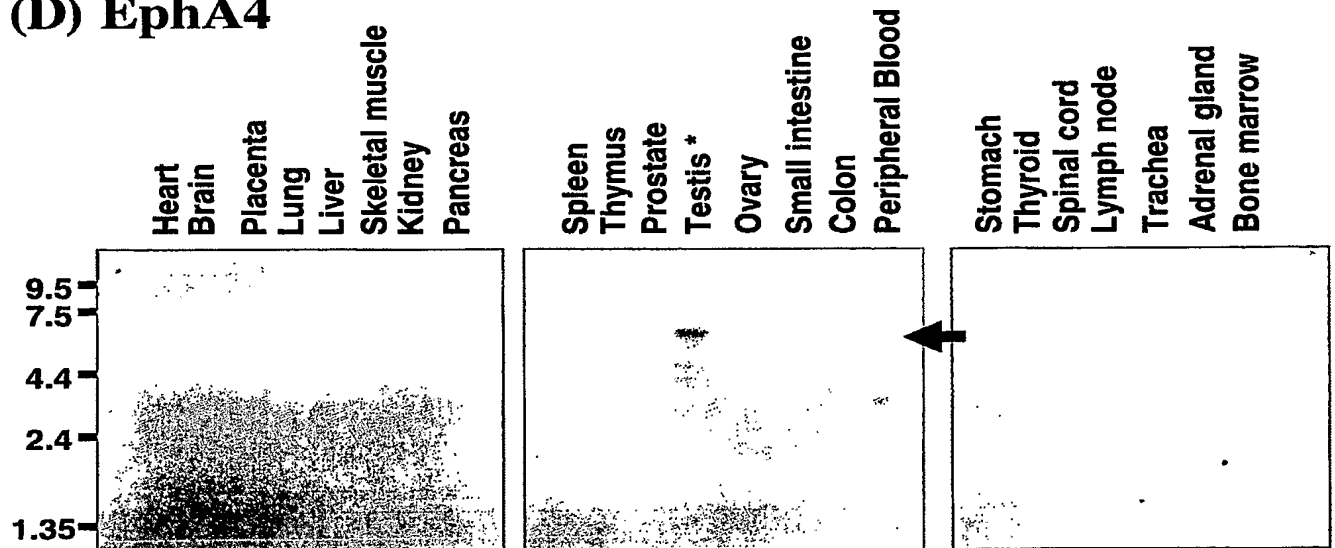
**(C) GPR107****(D) EphA4**

Fig. 4

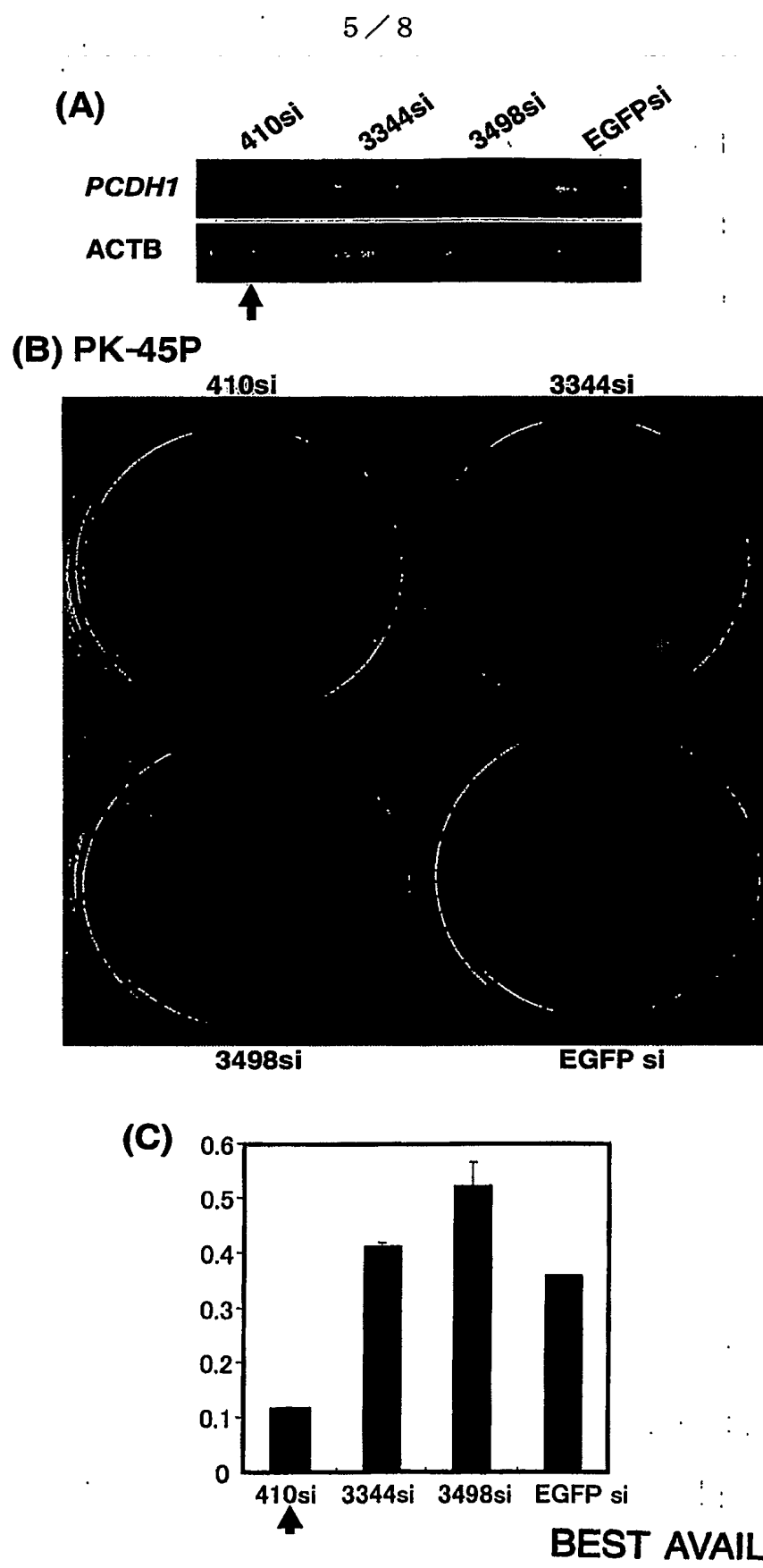




Fig. 5

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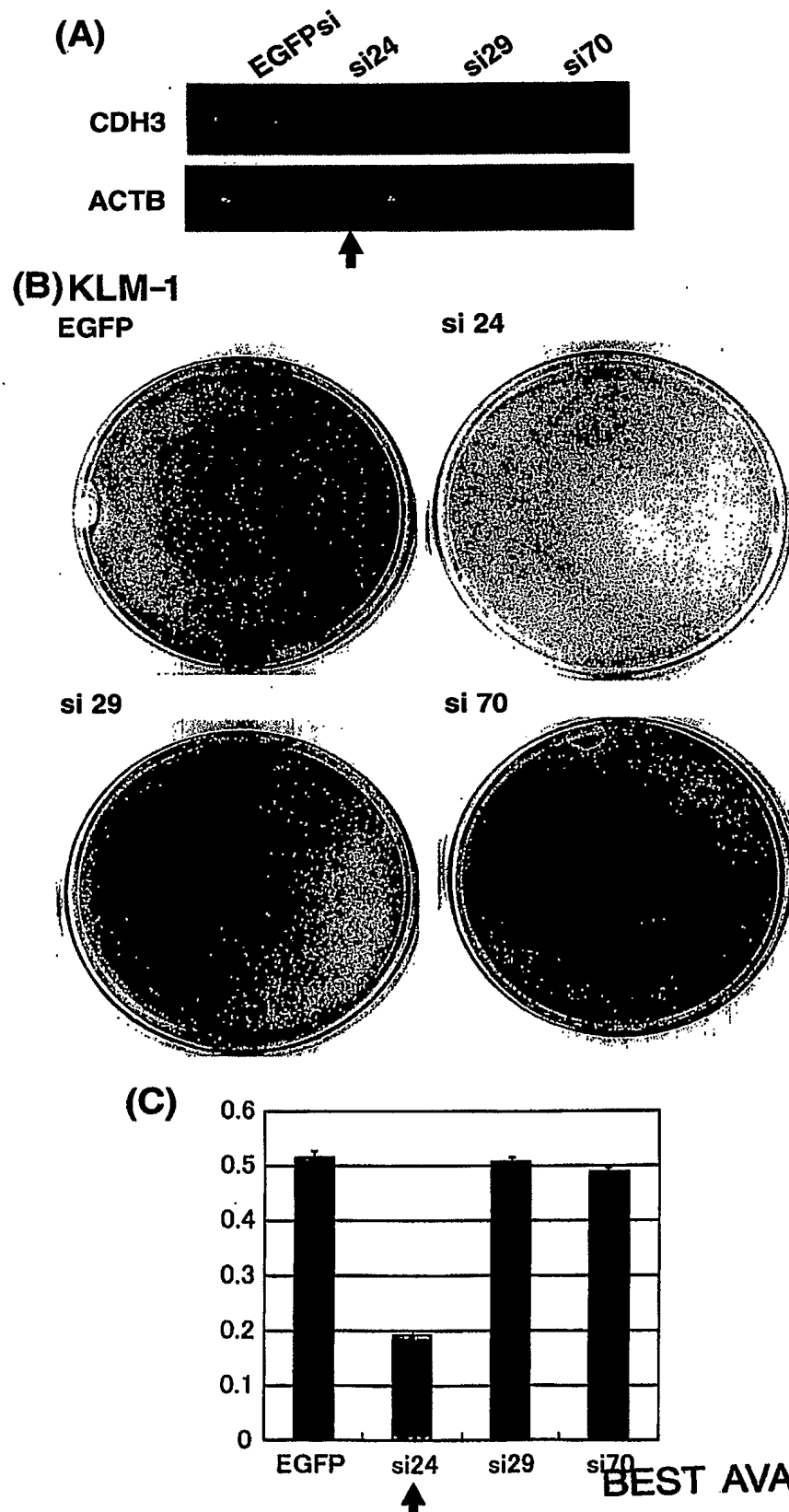
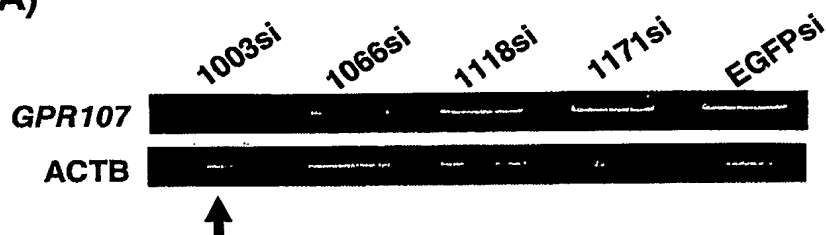


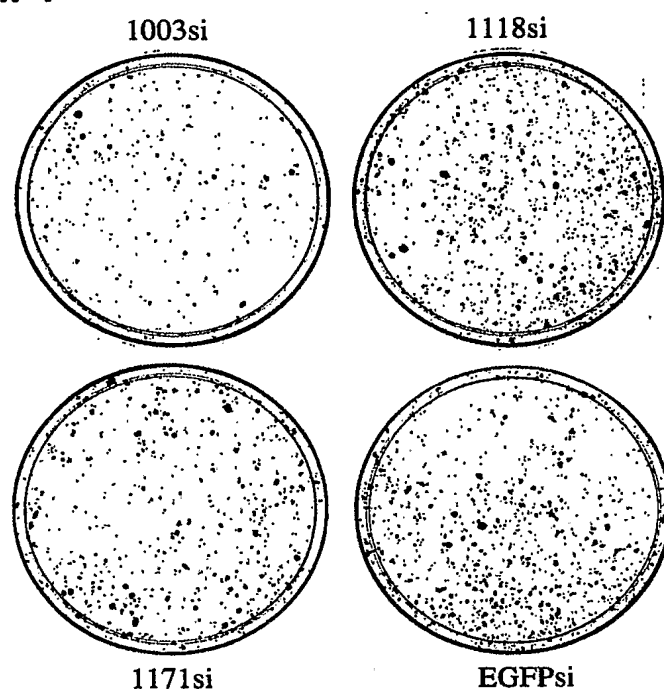
Fig. 6

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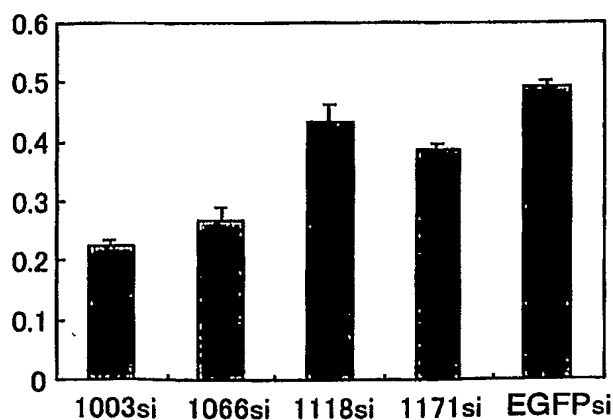
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(B) KLM-1



(C)

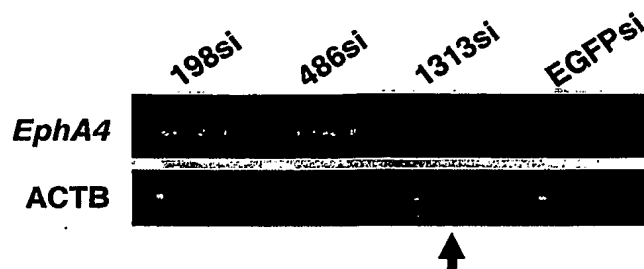


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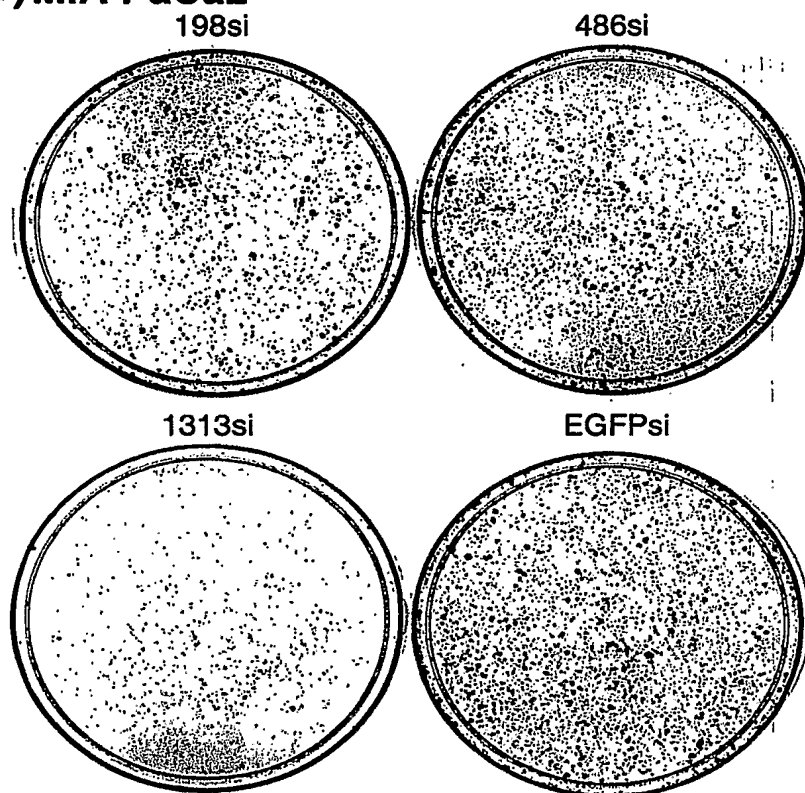
Fig. 8

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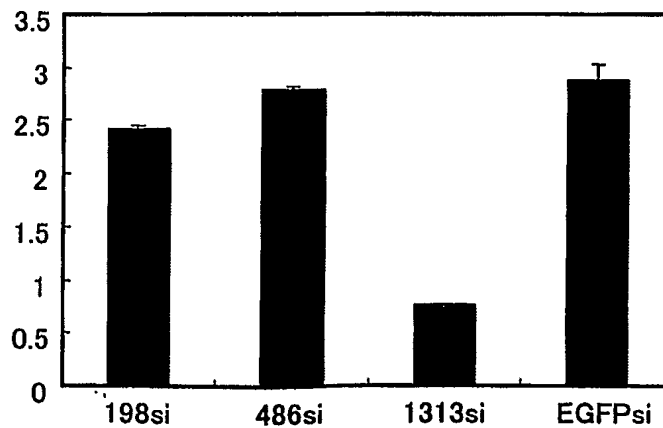
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(B) MIA-PaCa2



(C)



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Pro Ser Lys Arg Ile Leu Arg Arg His Lys Arg Asp Trp Val Val Ala  
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 50 55 60  
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 Phe Met Ile Val Ile Pro Leu Gln Val Leu Ala Asn Val Ala Tyr Ile  
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 Ile Ile Glu Ser Thr Glu Glu Gly Thr Thr Glu Tyr Gly Leu Trp Lys  
 385 390 395 400  
 Asp Ser Leu Phe Leu Val Asp Leu Leu Cys Cys Gly Ala Ile Leu Phe  
 405 410 415  
 Pro Val Val Trp Ser Ile Arg His Leu Gln Glu Ala Ser Ala Thr Asp  
 420 425 430  
 Gly Lys Gly Asp Ser Met Gly Pro Leu Gln Gln Arg Ala Asn Leu Arg  
 435 440 445  
 Ala Gly Ser Arg Ile Glu Ser Arg His Phe Ala Arg Ala Asp Leu Glu  
 450 455 460  
 Leu Leu Ala Ser Ser Cys Pro Pro Ala Ser Val Ser Gln Arg Ala Gly  
 465 470 475 480  
 Ile Thr Ala Ala Ile Asn Leu Ala Lys Leu Lys Leu Phe Arg His Tyr  
 485 490 495



Tyr Val Leu Ile Val Cys Tyr Ile Tyr Phe Thr Arg Ile Ile Ala Phe  
500 505 510  
Leu Leu Lys Leu Ala Val Pro Phe Gln Trp Lys Trp Leu Tyr Gln Leu  
515 520 525  
Leu Asp Glu Thr Ala Thr Leu Val Phe Phe Val Leu Thr Gly Tyr Lys  
530 535 540  
Phe Arg Pro Ala Ser Asp Asn Pro Tyr Leu Gln Leu Ser Gln Glu Glu  
545 550 555 560  
Glu Asp Leu Glu Met Glu Ser Val Val Thr Thr Ser Gly Val Met Glu  
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Ser Met Lys Lys Val Lys Lys Val Thr Asn Gly Ser Val Glu Pro Gln  
580 585 590  
Gly Glu Trp Glu Gly Ala Val  
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<212> DNA  
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Met Ala Gly Ile  
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ttc tat ttc gcc cta ttt tgc tgt ctc ttc ggg att tgc gac gct gtc 102  
Phe Tyr Phe Ala Leu Phe Ser Cys Leu Phe Gly Ile Cys Asp Ala Val  
5 10 15 20  
aca ggt tcc agg gta tac ccc gcg aat gaa gtt acc tta ttg gat tcc 150  
Thr Gly Ser Arg Val Tyr Pro Ala Asn Glu Val Thr Leu Leu Asp Ser  
25 30 35  
aga tct gtt cag gga gaa ctt ggg tgg ata gca agc cct ctg gaa gga 198  
Arg Ser Val Gln Gly Glu Leu Gly Trp Ile Ala Ser Pro Leu Glu Gly  
40 45 50  
ggg tgg gag gaa gtg agt atc atg gat gaa aaa aat aca cca atc cga 246  
Gly Trp Glu Glu Val Ser Ile Met Asp Glu Lys Asn Thr Pro Ile Arg  
55 60 65  
acc tac caa gtg tgc aat gtg atg gaa ccc agc cag aat aac tgg cta 294  
Thr Tyr Gln Val Cys Asn Val Met Glu Pro Ser Gln Asn Asn Trp Leu  
70 75 80  
cga act gat tgg atc acc cga gaa ggg got cag agg gtg tat att gag 342  
Arg Thr Asp Trp Ile Thr Arg Glu Gly Ala Gln Arg Val Tyr Ile Glu

85	90	95	100	
att aaa ttc acc ttg agg gac tgc aat agt ctt ccg ggc gtc atg ggg Ile Lys Phe Thr Leu Arg Asp Cys Asn Ser Leu Pro Gly Val Met Gly 105 110 115				390
act tgc aag gag acg ttt aac ctg tac tac tat gaa tca gac aac gac Thr Cys Lys Glu Thr Phe Asn Leu Tyr Tyr Tyr Glu Ser Asp Asn Asp 120 125 130				438
aaa gag cgt ttc atc aga gag aac cag ttt gtc aaa att gac acc att Lys Glu Arg Phe Ile Arg Glu Asn Gln Phe Val Lys Ile Asp Thr Ile 135 140 145				486
gct gct gat gag agc ttc acc caa gtg gac att ggt gac aga atc atg Ala Ala Asp Glu Ser Phe Thr Gln Val Asp Ile Gly Asp Arg Ile Met 150 155 160				534
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cta tgc aac gct ggg cat gag gag cgg agc gga gaa tgc caa gct tgc Leu Cys Asn Ala Gly His Glu Glu Arg Ser Gly Glu Cys Gln Ala Cys 265 270 275				870
aaa att gga tat tac aag gct ctc tcc acg gat gcc acc tgt gcc aag Lys Ile Gly Tyr Tyr Lys Ala Leu Ser Thr Asp Ala Thr Cys Ala Lys 280 285 290				918
tgc cca ccc cac agc tac tot gtc tgg gaa gga gcc acc tcg tgc acc Cys Pro Pro His Ser Tyr Ser Val Trp Glu Gly Ala Thr Ser Cys Thr 295 300 305				966
tgt gac cga ggc ttt ttc aga gct gac aac gat gct gcc tct atg ccc Cys Asp Arg Gly Phe Phe Arg Ala Asp Asn Asp Ala Ala Ser Met Pro 310 315 320				1014
tgc acc cgt cca cca tct gct ccc ctg aac ttg att tca aat gtc aac Cys Thr Arg Pro Pro Ser Ala Pro Leu Asn Leu Ile Ser Asn Val Asn				1062

325	330	335	340	
gag aca tct gtg aac ttg gaa tgg agt ago cct cag aat aca ggt ggc Glu Thr Ser Val Asn Leu Glu Trp Ser Ser Pro Gln Asn Thr Gly Gly 345 350 355				1110
cgc cag gac att tcc tat aat gtg gta tgc aag aaa tgt gga gct ggt Arg Gln Asp Ile Ser Tyr Asn Val Val Cys Lys Lys Cys Gly Ala Gly 360 365 370				1158
gac ccc agc aag tgc cga ccc tgt gga agt ggg gtc cac tac acc cca Asp Pro Ser Lys Cys Arg Pro Cys Gly Ser Gly Val His Tyr Thr Pro 375 380 385				1206
cag cag aat ggc ttg aag acc acc aaa gtc tcc atc act gac ctc cta Gln Gln Asn Gly Leu Lys Thr Thr Lys Val Ser Ile Thr Asp Leu Leu 390 395 400				1254
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aac aca gtg cct tcc cgg atc att gga gat ggg gct aac tcc aca gtc Asn Thr Val Pro Ser Arg Ile Ile Gly Asp Gly Ala Asn Ser Thr Val 535 540 545				1686
ott ctg gtc tot gtc tcc ggc agt gtg gtg ctg gtg gta att ctc att Leu Leu Val Ser Val Ser Gly Ser Val Val Leu Val Val Ile Leu Ile 550 555 560				1734
gca gct ttt gtc atc ago cgg aga cgg agt aaa tac agt aaa gcc aaa Ala Ala Phe Val Ile Ser Arg Arg Arg Ser Lys Tyr Ser Lys Ala Lys				1782

565	570	575	580	
caa gaa gcg gat Gln Glu Ala Asp	gaa gag aaa cat Glu Lys His	ttg aat caa ggt Leu Asn Gln Gly	gta aga aca tat Val Arg Thr Tyr	1830
	585	590	595	
gtg gac ccc ttt Val Asp Pro	acg tac gaa gat Thr Tyr Glu Asp	ccc aac caa goa Pro Asn Gln Ala	gtg cga gag ttt Val Arg Glu Phe	1878
	600	605	610	
gcc aaa gaa att Ala Lys Glu	gac goa tcc tgc Asp Ala Ser Cys	att aag att gaa Ile Lys Ile Glu	aaa gtt ata gga Lys Val Ile Gly	1926
	615	620	625	
gtt ggt gaa ttt Val Gly Glu	gtt ggt gag gta Gly Glu Val	tgc agt ggg cgt Cys Ser Gly Arg	ctc aaa gtg cct Leu Lys Val Pro	1974
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aag aga gag atc Lys Arg Glu	tgt gtt gct atc Cys Val Ala Ile	aag act ctg aaa Lys Thr Leu Lys	gct ggt tat aca Ala Gly Tyr Thr	2022
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gac aaa cag agg Asp Lys Gln Arg	aga gac ttc ctg Arg Asp Phe Leu	agt gag gcc agc Ser Glu Ala Ser	atc atg gga cag Ile Met Gly Gln	2070
	665	670	675	
ttt gac cat ccg Phe Asp His	aac atc att cac Asn Ile Ile His	ttg gaa ggc gtg Leu Glu Gly Val	gtc act aaa tgt Val Thr Lys Cys	2118
	680	685	690	
aaa cca gta atg Lys Pro Val	atc ata aca gag Ile Ile Thr Glu	atg gag aat ggc Met Glu Asn Gly	tcc ttg gat Ser Leu Asp	2166
	695	700	705	
gca ttc ctc agg Ala Phe Leu	aaa aat gat ggc Lys Asn Asp Gly	aga ttt aca gtc Arg Phe Thr Val	att cag ctg gtg Ile Gln Leu Val	2214
	710	715	720	
ggc atg ctt cgt Gly Met Leu	ggc att ggg tct Gly Ile Gly Ser	atg aag tat tta Met Lys Tyr Leu	tct gat atg Ser Asp Met	2262
	730	735	740	
agc tat gtg cat Ser Tyr Val	cgt gat ctg gcc Arg Asp Leu Ala	gca cgg aac atc Ala Arg Asn Ile	ctg gtg aac ago Leu Val Asn Ser	2310
	745	750	755	
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	760	765	770	
gat gat ccg gaa Asp Asp Pro	gca goa gct tac Ala Ala Tyr Thr	acc acc agg ggt Thr Thr Arg Gly	ggc aag att cct Gly Lys Ile Pro	2406
	775	780	785	
cgg tgg act gcg Arg Trp Thr	cca gaa gca att Pro Glu Ala Ile	gcc tat cgt aaa Ala Tyr Arg Lys	ttc aca tca gca Phe Thr Ser Ala	2454
	790	795	800	
agt gat gta tgg Ser Asp Val	ago tat gga atc Ser Tyr Gly Ile	gtt atg tgg gaa Val Met Trp Glu	gtg atg tog tac Val Met Ser Tyr	2502

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Gly Glu Arg Pro Tyr 825	Trp Asp Met Ser 830	Gln Asp Val Ile 835	Lys Ala	
att gag gaa ggc tat cgg tta ccc cct cca atg gac tgc ccc att ggc	2598			
Ile Glu Glu Gly Tyr 840	Arg Leu Pro Pro 845	Met Asp Cys Pro 850	Ile Ala	
ctc cac cag ctg atg cta gac tgc tgg cag aag gag agg agc gac agg	2646			
Leu His Gln Leu Met 855	Leu Asp Cys Trp 860	Gln Lys Glu Arg Ser 865	Asp Arg	
cct aaa ttt ggg cag att gtc aac atg ttg gac aaa ctc atc cgc aac	2694			
Pro Lys Phe Gly Gln Ile 870	Val Asn Met Leu 875	Asp Lys Leu Ile 880	Arg Asn	
ccc aac agc ttg aag agg aca ggg acg gag agc tcc aga cct aac act	2742			
Pro Asn Ser Leu Lys 885	Arg Thr Gly Thr 890	Glu Ser Arg Pro 895	Asn Thr 900	
gcc ttg ttg gat cca agc tcc cct gaa ttc tct gct gtg gta tca gtg	2790			
Ala Leu Leu Asp Pro 905	Ser Ser Pro Glu Phe 910	Ser Ala Val Val 915	Ser Val	
ggc gat tgg ctc cag gcc att aaa atg gac cgg tat aag gat aac ttc	2838			
Gly Asp Trp Leu Gln Ala Ile 920	Lys Met Asp Arg Tyr 925	Lys Asp Asn Phe 930		
aca gct gct ggt tat acc aca cta gag got gtg gtg cac gtg aac cag	2886			
Thr Ala Ala Gly Tyr Thr Thr 935	Leu Glu Ala Val Val 940	His Val Asn Gln 945		
gag gac ctg gca aga att ggt atc aca gcc atc acg cac cag aat aag	2934			
Glu Asp Leu Ala Arg Ile 950	Gly Ile Thr Ala Ile 955	Thr His Gln Asn Lys 960		
att ttg agc agt gtc cag gca atg cga acc caa atg cag cag atg cac	2982			
Ile Leu Ser Ser Val 965	Gln Ala Met Arg Thr 970	Gln Met Gln Gln Met 975	His 980	
ggc aga atg gtt ccc gtc tga gccagtactg aataaactca aaactcttga	3033			
Gly Arg Met Val Pro Val 985				
aattagttta cctcatccat gcactttaat tgaagaactg cacttttttt acttgcgttt	3093			
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35 40 45

Pro Leu Glu Gly Gly Trp Glu Glu Val Ser Ile Met Asp Glu Lys Asn  
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Thr Pro Ile Arg Thr Tyr Gln Val Cys Asn Val Met Glu Pro Ser Gln  
65 70 75 80

Asn Asn Trp Leu Arg Thr Asp Trp Ile Thr Arg Glu Gly Ala Gln Arg  
85 90 95

Val Tyr Ile Glu Ile Lys Phe Thr Leu Arg Asp Cys Asn Ser Leu Pro  
100 105 110

Gly Val Met Gly Thr Cys Lys Glu Thr Phe Asn Leu Tyr Tyr Tyr Glu  
115 120 125

Ser Asp Asn Asp Lys Glu Arg Phe Ile Arg Glu Asn Gln Phe Val Lys  
130 135 140

Ile Asp Thr Ile Ala Ala Asp Glu Ser Phe Thr Gln Val Asp Ile Gly  
145 150 155 160

Asp Arg Ile Met Lys Leu Asn Thr Glu Ile Arg Asp Val Gly Pro Leu  
165 170 175

Ser Lys Lys Gly Phe Tyr Leu Ala Phe Gln Asp Val Gly Ala Cys Ile  
180 185 190

Ala Leu Val Ser Val Arg Val Phe Tyr Lys Lys Cys Pro Leu Thr Val  
195 200 205

Arg Asn Leu Ala Gln Phe Pro Asp Thr Ile Thr Gly Ala Asp Thr Ser  
210 215 220

Ser Leu Val Glu Val Arg Gly Ser Cys Val Asn Asn Ser Glu Glu Lys  
225 230 235 240

Asp Val Pro Lys Met Tyr Cys Gly Ala Asp Gly Glu Trp Leu Val Pro  
245 250 255

Ile Gly Asn Cys Leu Cys Asn Ala Gly His Glu Glu Arg Ser Gly Glu  
260 265 270

Cys Gln Ala Cys Lys Ile Gly Tyr Tyr Lys Ala Leu Ser Thr Asp Ala  
275 280 285

Thr Cys Ala Lys Cys Pro Pro His Ser Tyr Ser Val Trp Glu Gly Ala  
290 295 300

Thr Ser Cys Thr Cys Asp Arg Gly Phe Phe Arg Ala Asp Asn Asp Ala

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Ala Ser Met Pro	Cys 325	Thr Arg Pro Pro	Ser 330	Ala Pro Leu Asn Leu Ile		335
Ser Asn Val	Asn 340	Glu Thr Ser Val	Asn 345	Leu Glu Trp Ser Ser Pro Gln		350
Asn Thr	Gly 355	Gly Arg Gln Asp Ile	Ser 360	Tyr Asn Val Val Cys Lys Lys		365
Cys Gly Ala Gly Asp Pro	Ser 375	Lys Cys Arg Pro	Cys 380	Gly Ser Gly Val		
His Tyr Thr Pro Gln Gln	Asn 390	Gly Leu Lys Thr Thr Lys Val Ser Ile				400
Thr Asp Leu Leu	Ala 405	His Thr Asn Tyr Thr Phe Glu Ile Trp Ala Val				415
Asn Gly Val Ser Lys Tyr Asn Pro	Asn 425	Pro Asp Gln Ser Val Ser Val				
Thr Val Thr Thr Asn Gln Ala Ala Pro Ser Ser Ile Ala Leu Val Gln						
	435		440		445	
Ala Lys Glu Val Thr Arg Tyr Ser Val Ala Leu Ala Trp Leu Glu Pro						
	450		455		460	
Asp Arg Pro Asn Gly Val Ile Leu Glu Tyr Glu Val Lys Tyr Tyr Glu						
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Lys Asp Gln Asn Glu Arg Ser Tyr Arg Ile Val Arg Thr Ala Ala Arg						
	485		490		495	
Asn Thr Asp Ile Lys Gly Leu Asn Pro Leu Thr Ser Tyr Val Phe His						
	500		505		510	
Val Arg Ala Arg Thr Ala Ala Gly Tyr Gly Asp Phe Ser Glu Pro Leu						
	515		520		525	
Glu Val Thr Thr Asn Thr Val Pro Ser Arg Ile Ile Gly Asp Gly Ala						
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Asn Ser Thr Val Leu Leu Val Ser Val Ser Gly Ser Val Val Leu Val						
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Val Ile Leu Ile Ala Ala Phe Val Ile Ser Arg Arg Arg Ser Lys Tyr						
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Ser Lys Ala Lys Gln Glu Ala Asp Glu Glu Lys His Leu Asn Gln Gly						
	580		585		590	
Val Arg Thr Tyr Val Asp Pro Phe Thr Tyr Glu Asp Pro Asn Gln Ala						
	595		600		605	
Val Arg Glu Phe Ala Lys Glu Ile Asp Ala Ser Cys Ile Lys Ile Glu						
	610		615		620	
Lys Val Ile Gly Val Gly Glu Phe Gly Glu Val Cys Ser Gly Arg Leu						

625		630		635		640
Lys Val Pro Gly	Lys Arg Glu Ile Cys	Val Ala Ile Lys Thr	Leu Lys			
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Ala Gly Tyr Thr	Asp Lys Gln Arg Arg	Asp Phe Leu Ser	Glu Ala Ser			
	660	665	670			
Ile Met Gly Gln	Phe Asp His Pro Asn	Ile Ile His	Leu Glu Gly Val			
	675	680	685			
Val Thr Lys Cys	Lys Pro Val Met Ile	Ile Thr Glu Tyr	Met Glu Asn			
	690	695	700			
Gly Ser Leu Asp	Ala Phe Leu Arg Lys	Asn Asp Gly Arg	Phe Thr Val			
705	710	715	720			
Ile Gln Leu Val	Gly Met Leu Arg Gly	Ile Gly Ser Gly	Met Lys Tyr			
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Leu Ser Asp Met	Ser Tyr Val His Arg	Asp Leu Ala Ala	Arg Asn Ile			
	740	745	750			
Leu Val Asn Ser	Asn Leu Val Cys Lys	Val Ser Asp Phe	Gly Met Ser			
	755	760	765			
Arg Val Leu Glu	Asp Asp Pro Glu Ala	Ala Tyr Thr Thr	Arg Gly Gly			
	770	775	780			
Lys Ile Pro Ile	Arg Trp Thr Ala Pro	Glu Ala Ile Ala	Tyr Arg Lys			
785	790	795	800			
Phe Thr Ser Ala	Ser Asp Val Trp Ser	Tyr Gly Ile Val	Met Trp Glu			
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Val Met Ser Tyr	Gly Glu Arg Pro Tyr	Trp Asp Met Ser	Asn Gln Asp			
	820	825	830			
Val Ile Lys Ala	Ile Glu Glu Gly Tyr	Arg Leu Pro Pro	Pro Met Asp			
	835	840	845			
Cys Pro Ile Ala	Leu His Gln Leu Met	Leu Asp Cys Trp	Gln Lys Glu			
	850	855	860			
Arg Ser Asp Arg	Pro Lys Phe Gly Gln	Ile Val Asn Met	Leu Asp Lys			
865	870	875	880			
Leu Ile Arg Asn	Pro Asn Ser Leu Lys	Arg Thr Gly Thr	Glu Ser Ser			
	885	890	895			
Arg Pro Asn Thr	Ala Leu Leu Asp Pro	Ser Ser Pro Glu	Phe Ser Ala			
	900	905	910			
Val Val Ser Val	Gly Asp Trp Leu Gln	Ala Ile Lys Met	Asp Arg Tyr			
	915	920	925			
Lys Asp Asn Phe	Thr Ala Ala Gly Tyr	Thr Thr Leu Glu	Ala Val Val			
	930	935	940			
His Val Asn Gln	Glu Asp Leu Ala Arg	Ile Gly Ile Thr	Ala Ile Thr			



945					950					955					960
His	Gln	Asn	Lys	Ile	Leu	Ser	Ser	Val	Gln	Ala	Met	Arg	Thr	Gln	Met
				965					970					975	
Gln	Gln	Met	His	Gly	Arg	Met	Val	Pro	Val						
			980					985							

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$\langle 211 \rangle$	23
$\langle 212 \rangle$	DNA
$\langle 213 \rangle$	Artificial

**<223> An artificially synthesized primer sequence for RT-PCR**

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$\langle 210 \rangle$	10
$\langle 211 \rangle$	24
$\langle 212 \rangle$	DNA
$\langle 213 \rangle$	Artificial

**<223> An artificially synthesized primer sequence for RT-PCR**

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$\langle 210 \rangle$	11
$\langle 211 \rangle$	21
$\langle 212 \rangle$	DNA
$\langle 213 \rangle$	Artificial

**<220>**  
**<223> An artificially synthesized primer sequence for RT-PCR**

<400> 11  
ctgaaggcgg ctaacacaga c 21

$\langle 210 \rangle$	12
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$\langle 212 \rangle$	DNA
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**<220>**  
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<213> Artificial

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23

<210> 14

<211> 23

<212> DNA

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<400> 14

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23

<210> 15

<211> 51

<212> DNA

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51

<210> 16

<211> 51

<212> DNA

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51

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<212> DNA

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51

<210> 18

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<212> DNA

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51

<210> 19

<211> 51

<212> DNA

<213> Artificial

<220>

<223> An artificially synthesized sequence for siRNA

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51

<210> 20

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51

<210> 21

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<212> DNA

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51

<210> 22

<211> 51

<212> DNA

<213> Artificial

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<400> 22

aaaaggagac aggtctggtt ttgtctcttg aacaacaacc agcctgtctc c

51

<210> 23

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<212> DNA

<213> Artificial

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51

<210> 24

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<210> 25

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<213> Artificial

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<210> 26

<211> 51

<212> DNA

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51

<210> 27

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gacatcaatg acaacacac

19

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attccgtccg gcttcagat

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gacttggaaa tggagtccg

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gaaagtcaag aaggtgacc.

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tccgaacctt ccaagtgtg

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